



Supramolecular Chemistry and Molecular Self-Assembly

M2 – Ecole polytechnique - Université Paris-Saclay
France

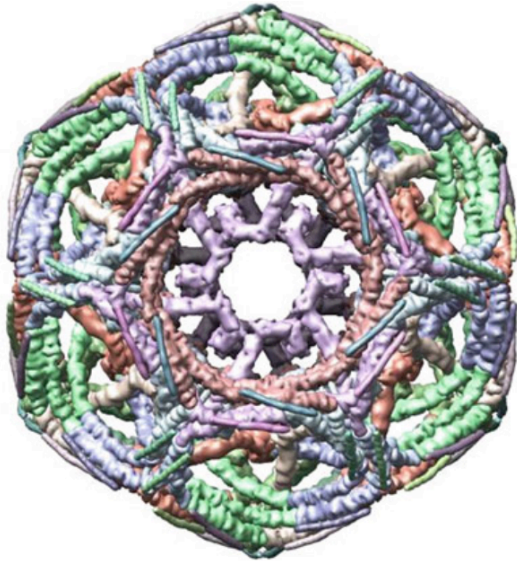
**Supramolecular chemistry in diagnosis and
therapeutic chemistry**

Outline

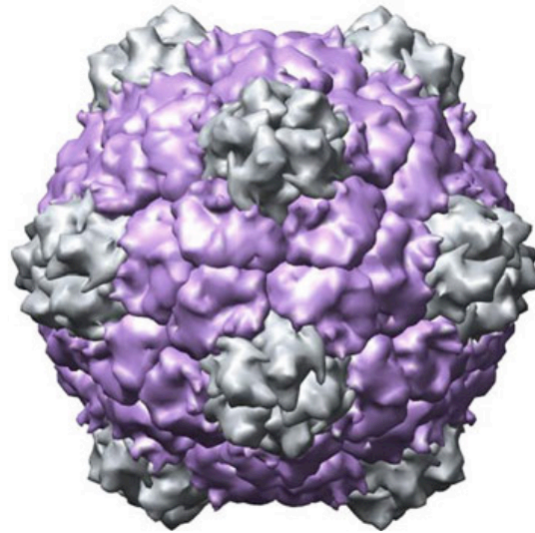
1. Thermodynamic and kinetic aspects
2. Calyxarenes, dendrimers and other examples
3. Supramolecular chemistry in diagnosis and therapeutic chemistry
4. Supramolecular chemistry in the environment
5. Discussion

Artificial cells

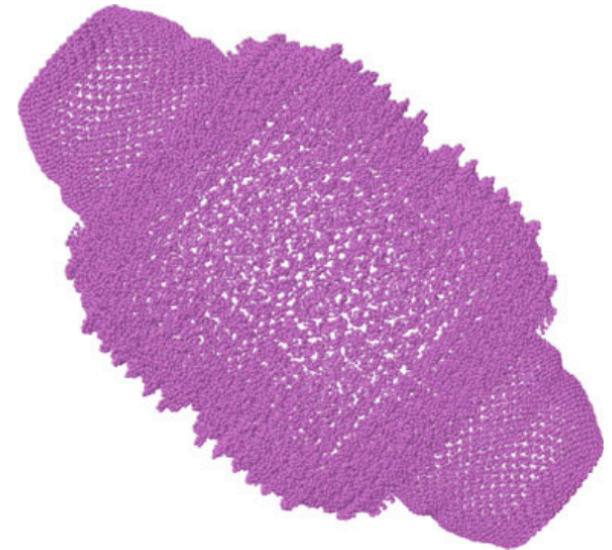
Nature is full of complex constructions designed to encapsulate molecules within a defined space.



A clathrin

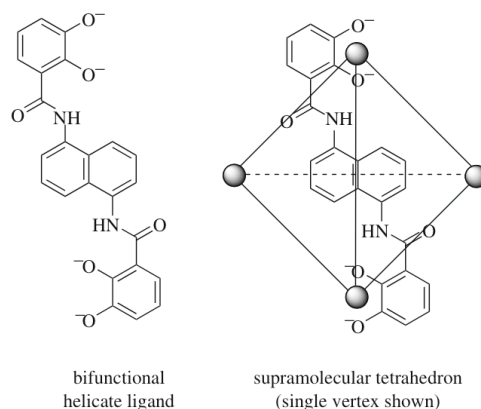


A capsid virus



A protein vault

Molecular cages: coordination chemistry



Raymond, *Angew. Chem. Int. Ed.*, 2006, 83

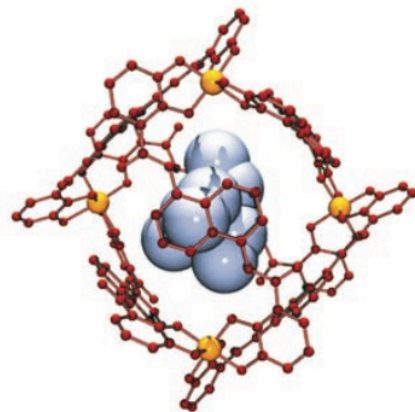
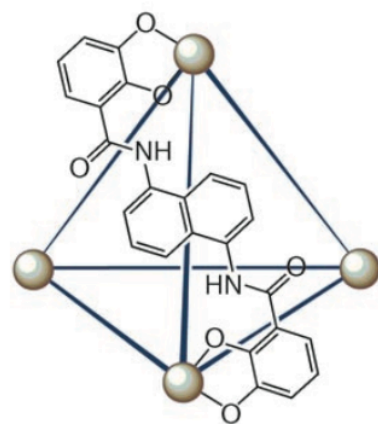
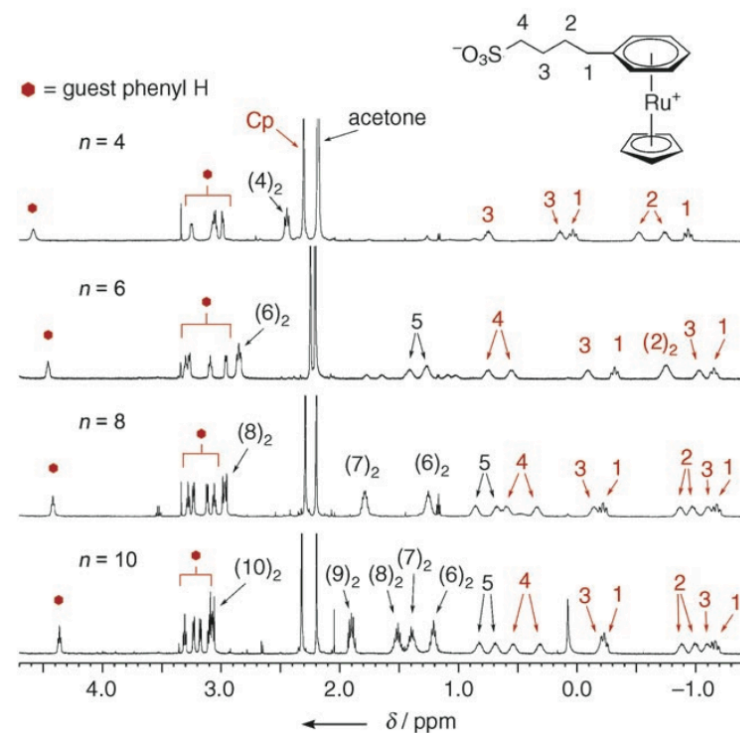


Figure 1. Left: Schematic structure of the M_4L_6 tetrahedral cluster which illustrates the structure of L^{4-} and its coordination to the metal-ion vertices. Right: Illustration of the host-guest complex $[Et_4N^+Ga_4L_6]^{11-}$ based on the X-ray structure coordinates, with a Et_4N^+ guest shown in blue.



Molecular cages: coordination chemistry

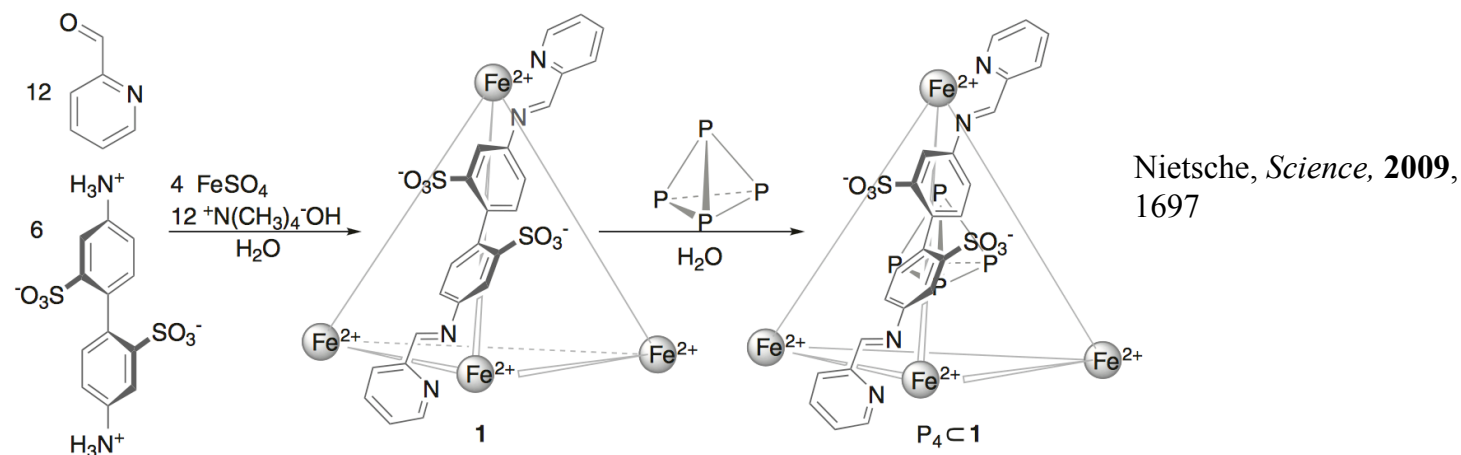


Fig. 1. Synthesis of tetrahedral cage **1** and subsequent incorporation of P_4 .

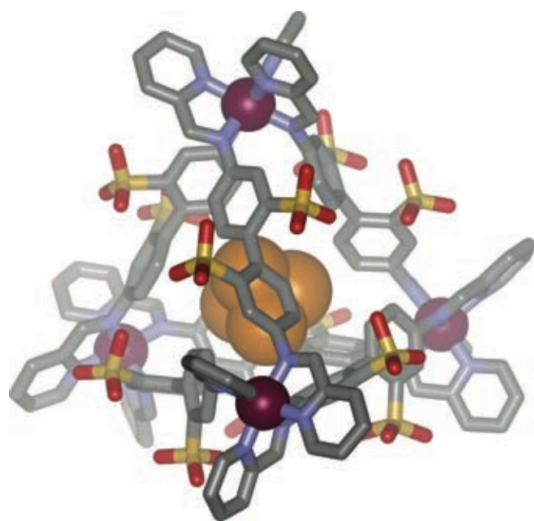
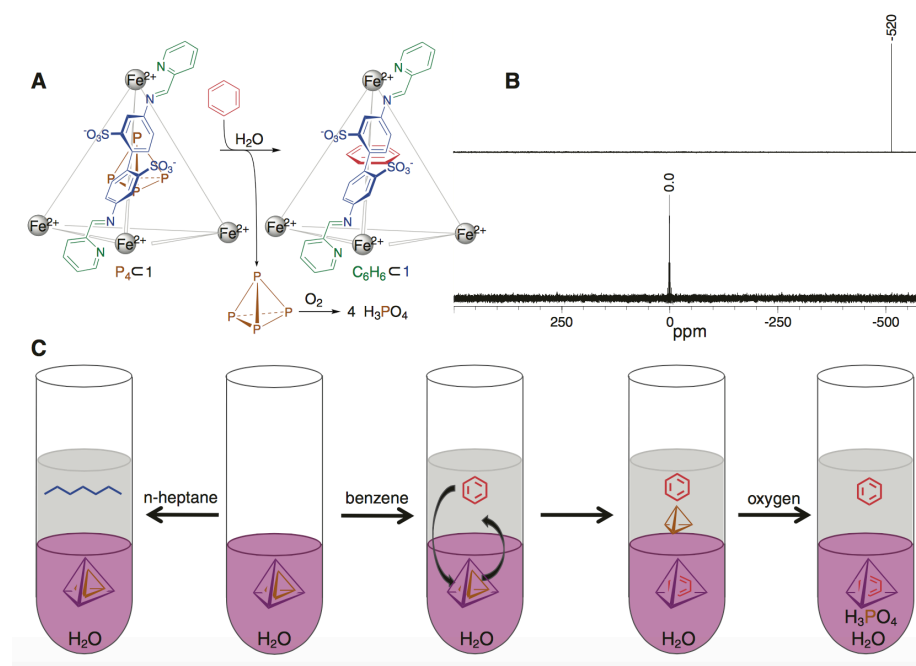


Fig. 2. Crystal structure of P_4C1 . Solvent molecules, counterions, and hydrogen atoms are omitted for clarity. Fe, violet; N, blue; C, gray; O, red; S, yellow; P, orange.



Supramolecular enzyme mimics: cytochrome models

Neumann, *J. Am. Chem. Soc.*,
1989, 111, 2900

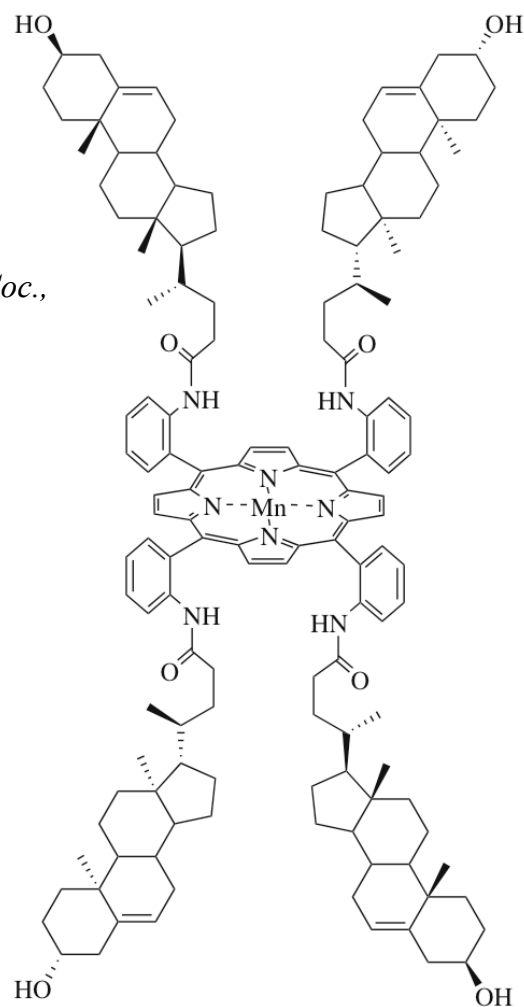


Fig. 4.10 Cytochrome model complexes [22, 23]

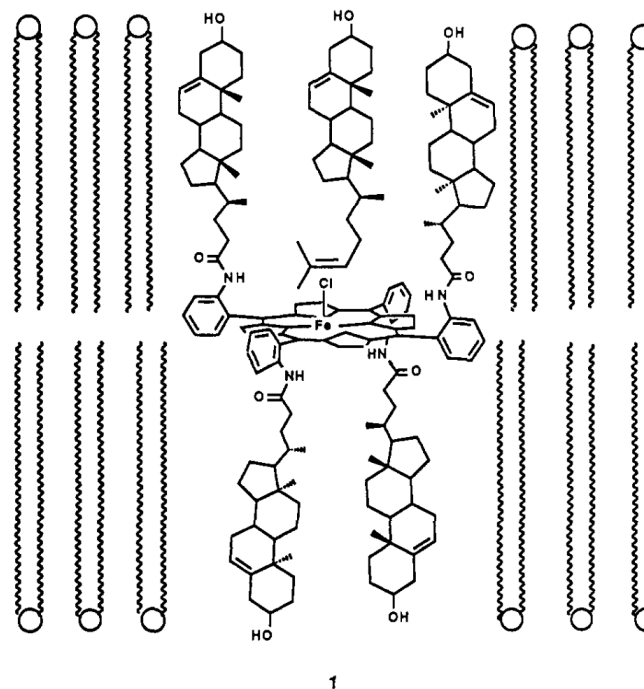
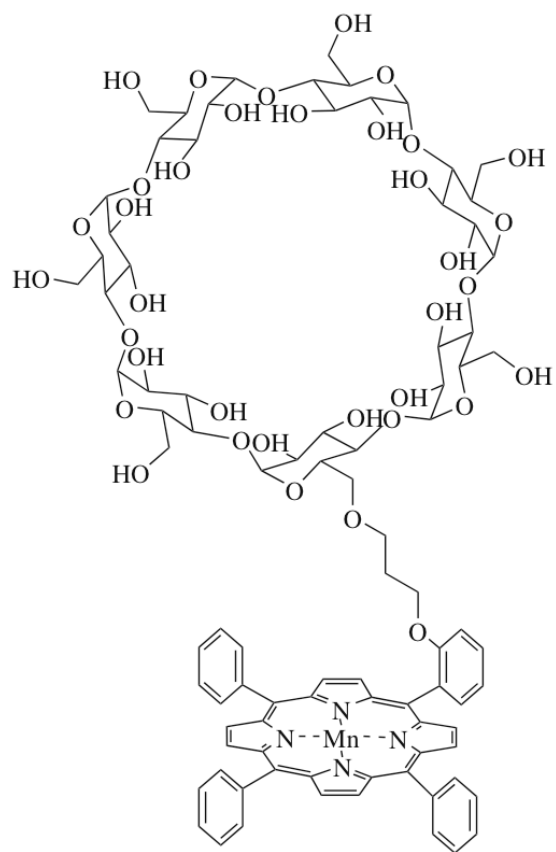


Figure 1. Idealized drawing of molecular bilayer assembly of chloro-[$\alpha,\beta,\alpha,\beta$ -*meso*-tetrakis[*o*-((3 β -hydroxy-5-cholestenyl)amido)phenyl]-porphyrinato]iron(III) ($\text{Fe}^{\text{III}}(\text{ChP})\text{Cl}$), desmosterol, and phospholipid.

When introduced to vesicles the complex buried itself in the membrane and, once flavoprotein pyruvate oxidase and ethylbenzene were added, generated acetophenone.

Supramolecular enzyme mimics: cytochrome models



Breslow, *Angew. Chem. Int. Ed.*, **2000**, *39*, 2692

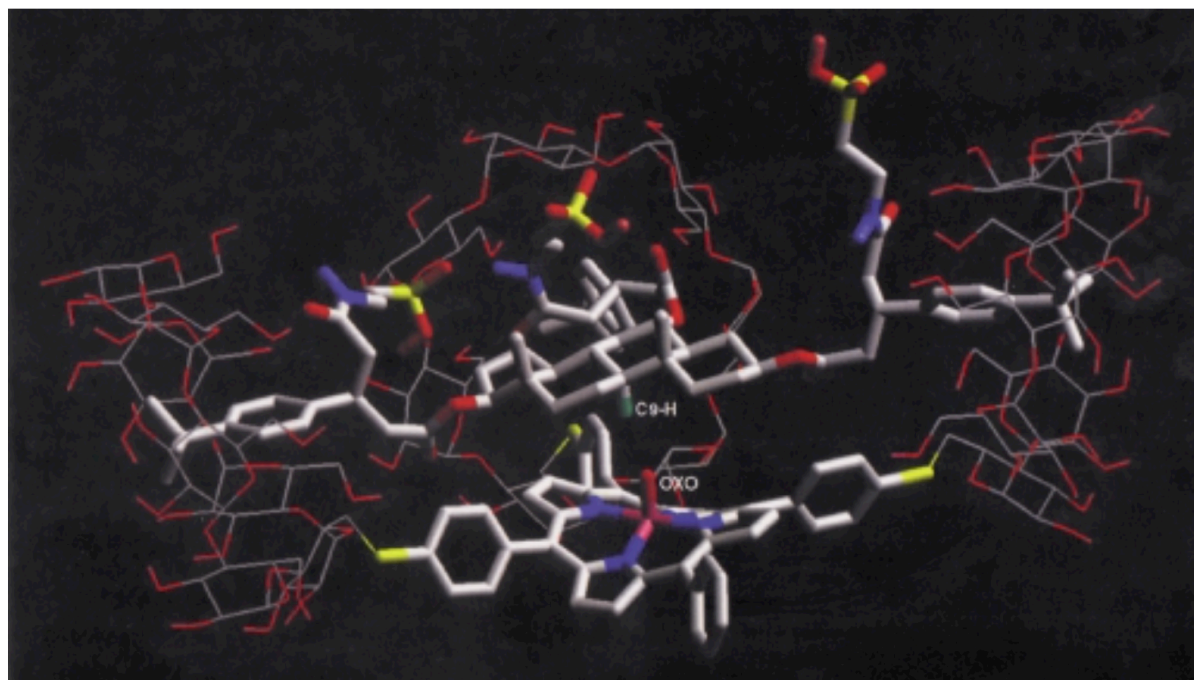
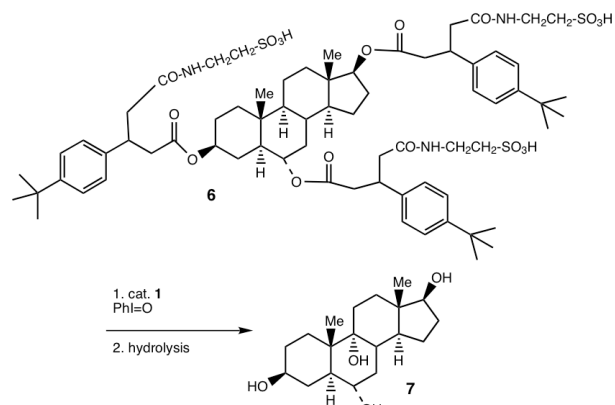


Figure 2. The computed structure of the complex of triply bound substrate **6** with catalyst **1**, showing that the oxygen atom added to the Mn atom of **1** is in a lateral position to attack the axial H atom on C-9 of **6**, as observed. For details of the calculational method, which differs from that in Figure 1, see Experimental Section. For clarity the one cyclodextrin not involved in substrate binding is deleted from the figure, and those binding the substrate are shown with thin lines.



Scheme 2. Catalyst **1** converts a triply bound substrate to the 9 α -hydroxy derivative.

Supramolecular enzyme mimics: cytochrome c oxidase

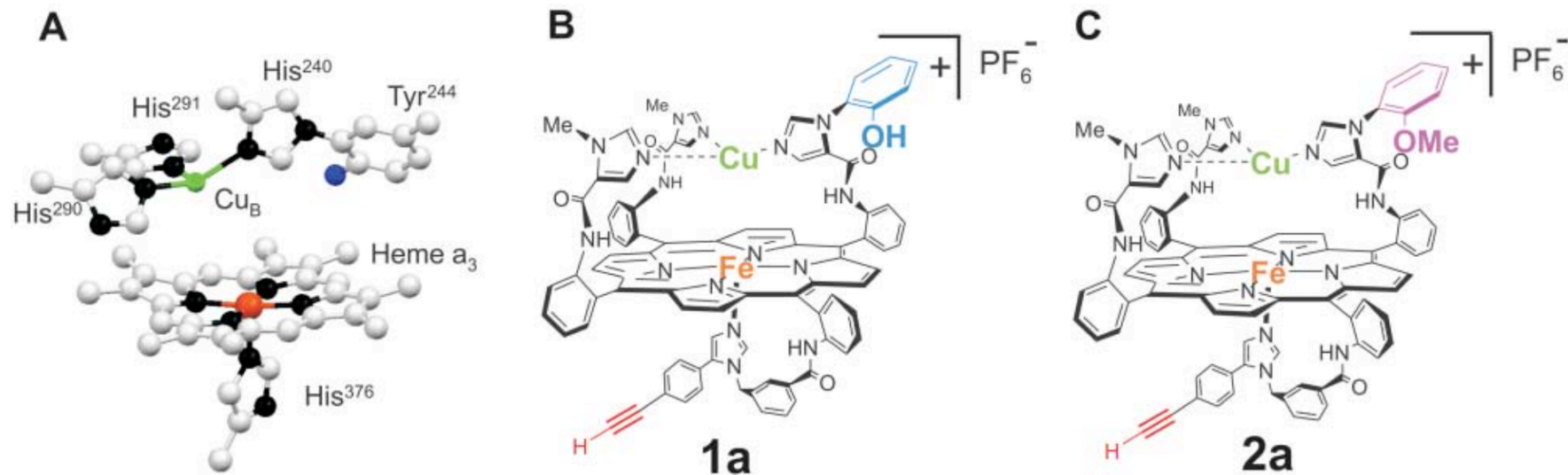
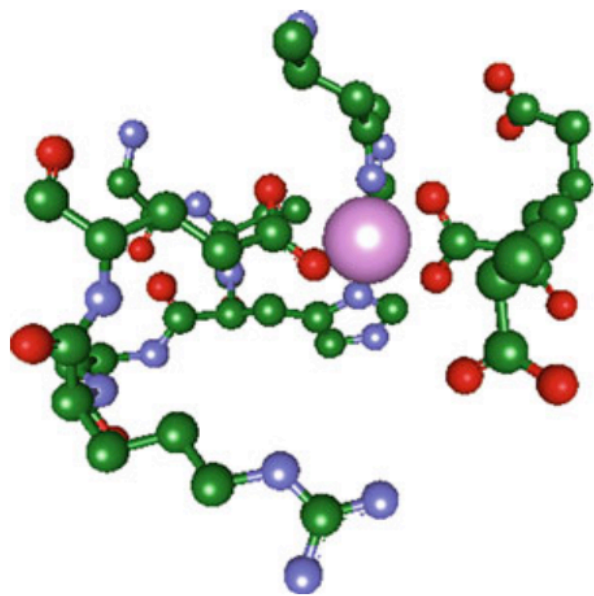


Fig. 1. (A) Crystal structure of the active site of CcO from the bovine heart (13). (B) Model **1a** reproduces the key elements of the active site of CcO. (C) Model **2a**, in which the phenol is masked as a methyl ether. Model **2a** can be treated with dilute acid to yield the iron-only model (**2b**), which is not shown.

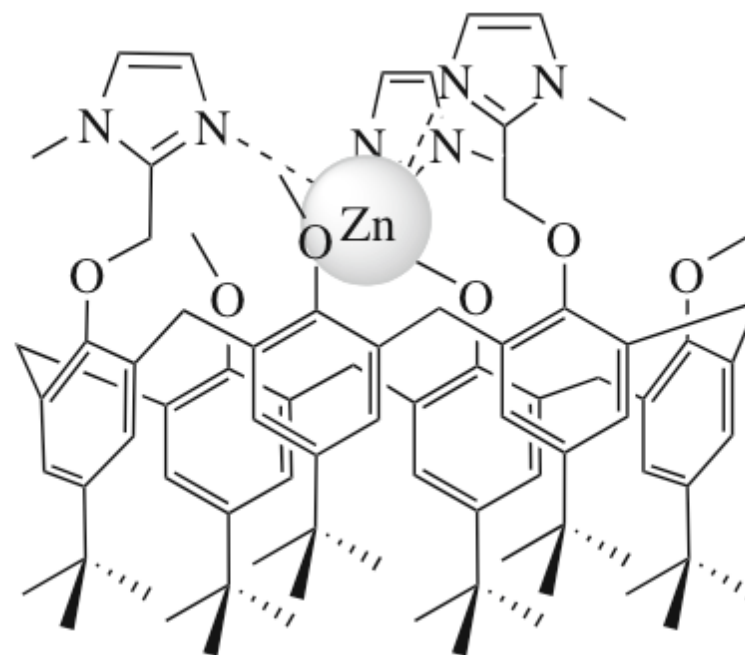
Cytochrome c oxidase mimics have long been a goal because they catalyse the reduction of dioxygen to water without the generation of other reactive oxygen species.

X-Ray crystallography revealed that the active site consisted of an iron-haem complex with an adjacent copper cation bound by three histidines, in the expected motif, and a nearby tyrosine group. The importance of tyrosine had been overlooked in many model systems but was introduced as an imidazole substituent by the Collman group

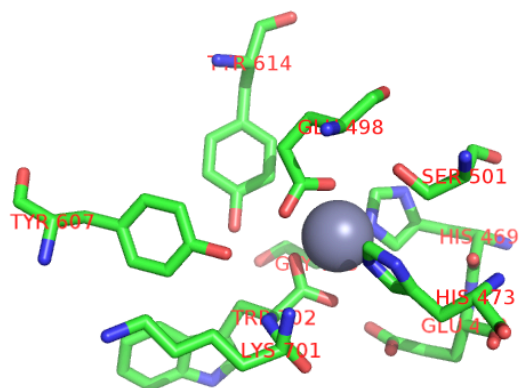
Supramolecular enzyme mimics: zinc carboxypeptidase



The zinc binding site in carboxypeptidase



Reinaud, O. *Angew Chem Int Ed Engl*, **2002**, *41*, 1044



Supramolecular enzyme mimics: carbonic anhydrase

Synthesis and structure of $[\text{Zn}(\text{OMe})(\text{L})]\cdot[\text{Zn}(\text{OH})(\text{L})]\cdot 2(\text{BPh}_4)$, $\text{L} = \text{cis,cis-1,3,5-tris}[(E,E)\text{-3-(2-furyl)acrylideneamino}]$ cyclohexane: structural models of carbonic anhydrase and liver alcohol dehydrogenase†

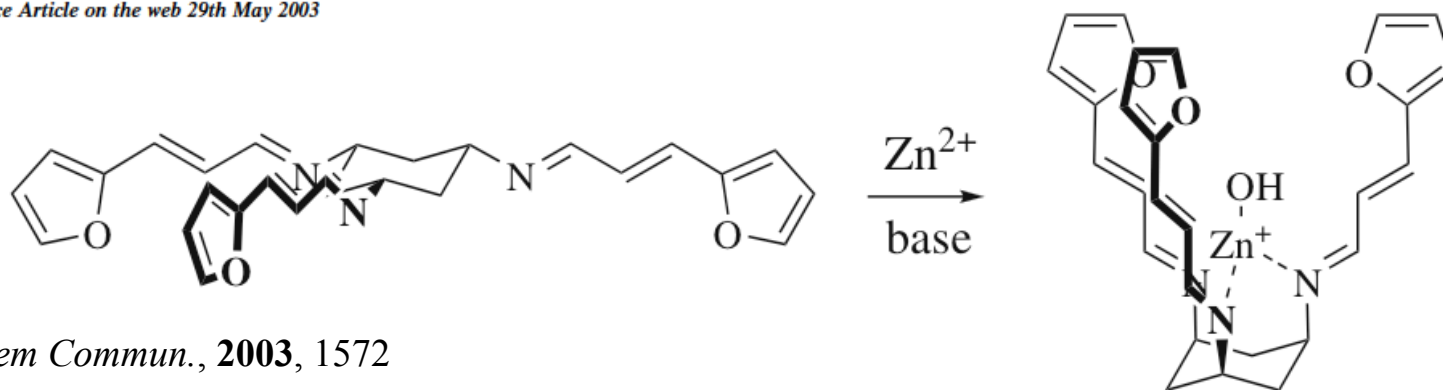
Leroy Cronin‡ and Paul H. Walton*

Department of Chemistry, The University of York, Heslington, York, UK YO10 5DD.

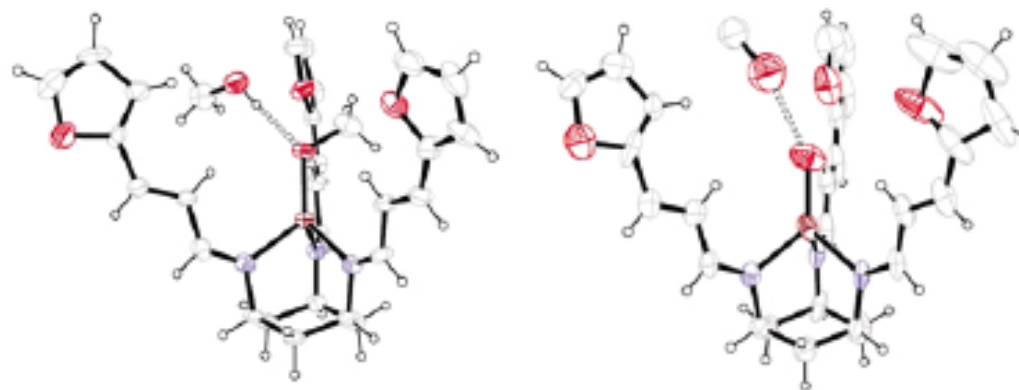
E-mail: phw2@york.ac.uk; Fax: +44 1904 432516; Tel: +44 1904 432580

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First published as an Advance Article on the web 29th May 2003



Walton, *Chem Commun.*, **2003**, 1572



Supramolecular enzyme mimics: ATPase

A crown ether ATPase

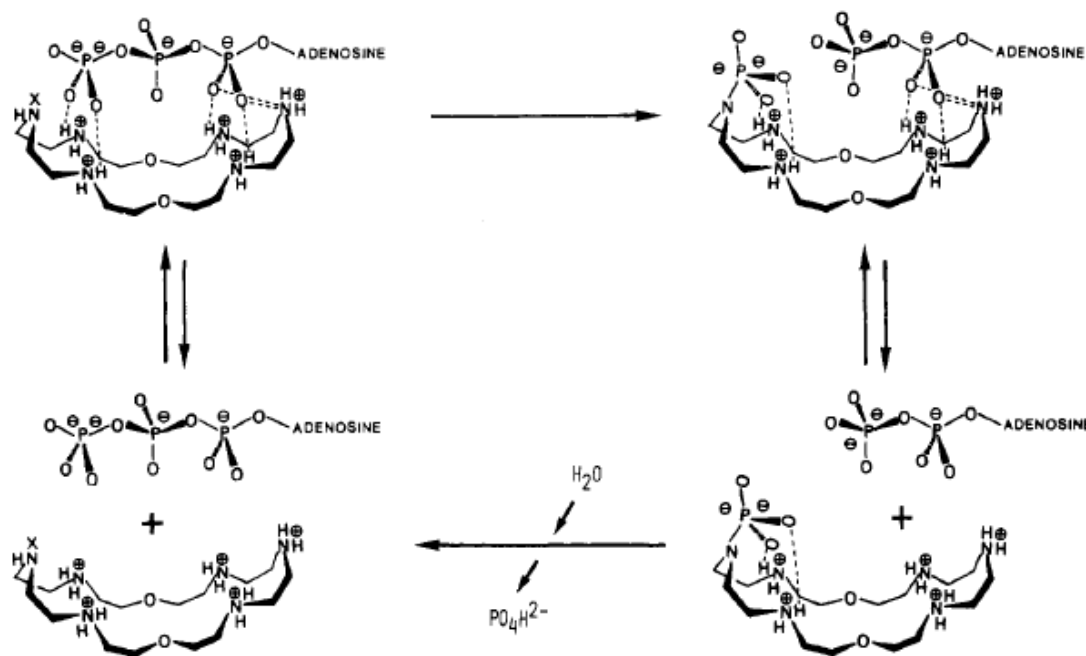
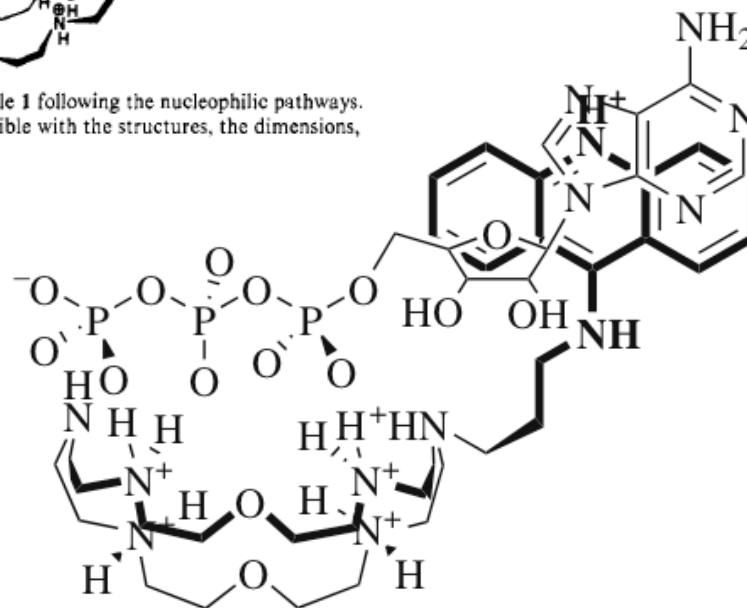
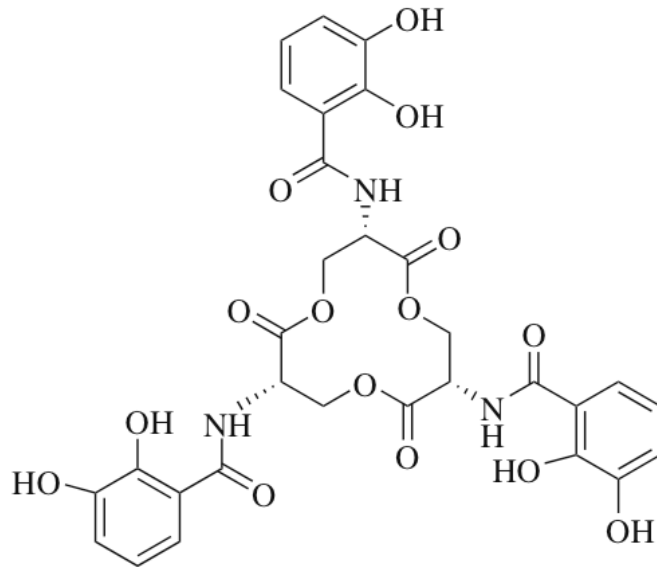


Figure 3. Schematic representation of the catalytic cycle for ATP hydrolysis by the macrocyclic receptor molecule **1** following the nucleophilic pathways. The geometry of the ATP·**1** complex and the binding scheme are hypothetical (see text and ref 11) but compatible with the structures, the dimensions, and the binding-site arrangement of the two partners.

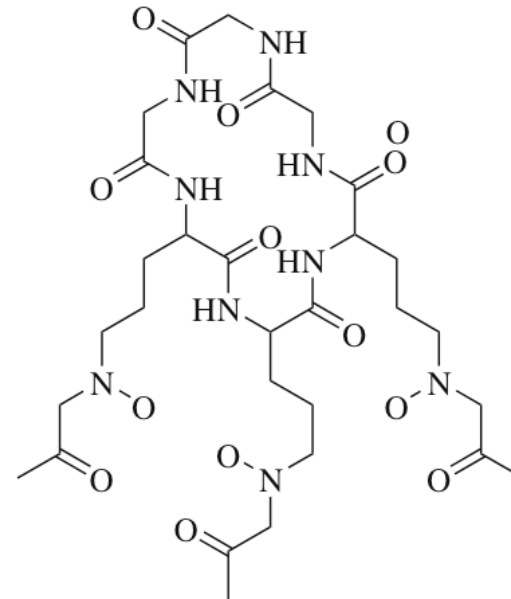


Supramolecular enzyme mimics: Siderophores

Yeast, fungi and bacteria, such as the *Salmonella* genus and *Escherichia coli*, need to sequester iron from aqueous solution. Unfortunately water soluble Fe^{2+} is easily oxidized to Fe^{3+} in air which makes it much less soluble. At concentrations of around 10^{-17} M for free Fe^{3+} in aqueous solution it is essential that every available ion is bound effectively and transported across the cell membrane. This is achieved by siderophores, iron-specific chelating agents that bind Fe^{3+} in an octahedral pocket usually comprising three bidentate catechol or hydroxamate groups



enterobactin



ferrichrome

Supramolecular enzyme mimics: Anion Transport

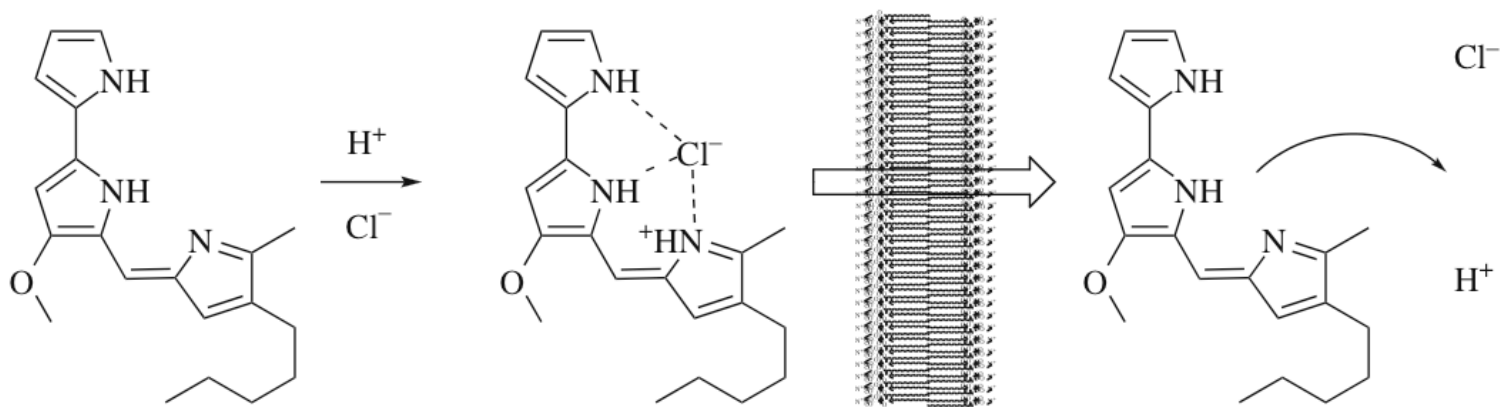


Fig. 5.4 Proton and chloride co-transport by prodigiosin

Anions can also be transported across membranes. In Nature, Cl^- is transported by prodigiosins which comprise three conjugated pyrrole groups and a lipophilic alkyl tail

Sato T et al., *J Biol Chem*, **1998**, 273, 21455

Transmembrane Channels: Selectivity and Gating Mechanisms

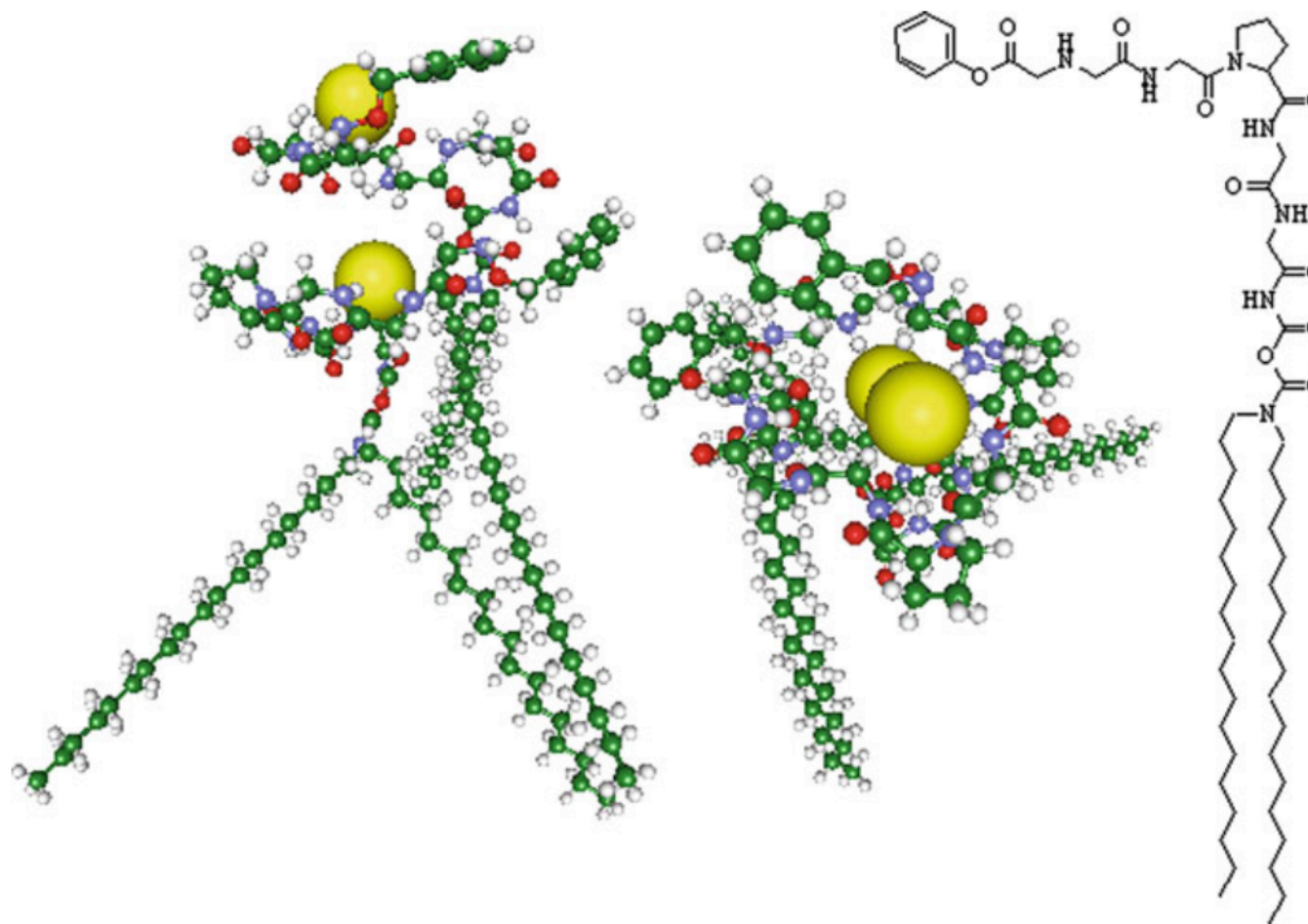


Fig. 5.6 Chloride transport by peptide aggregation in an artificial system [14]

Ion Transport Ionophores and Siderophores

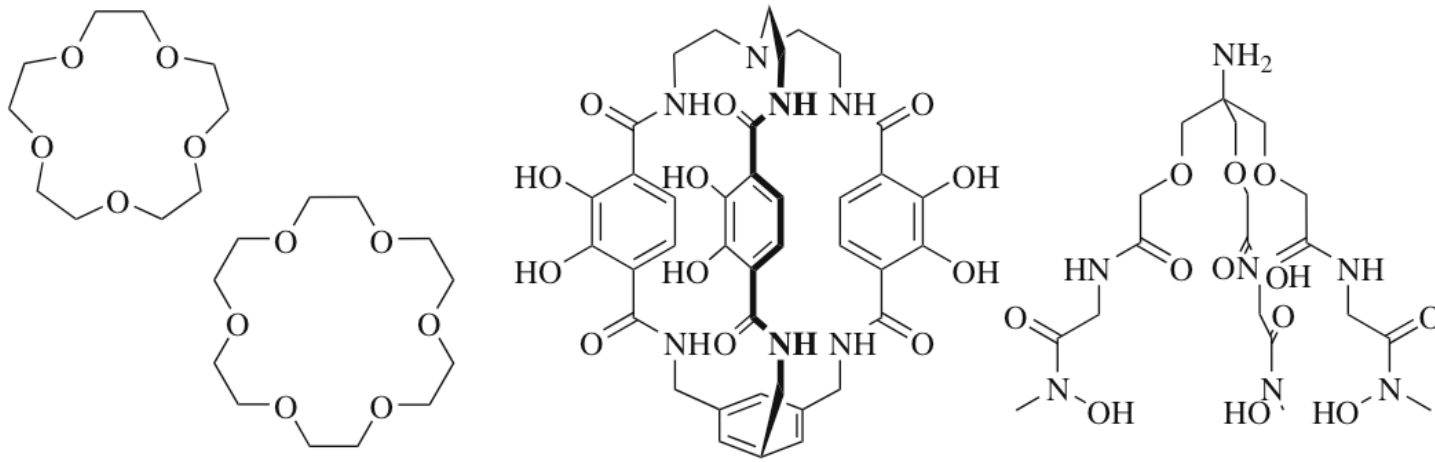
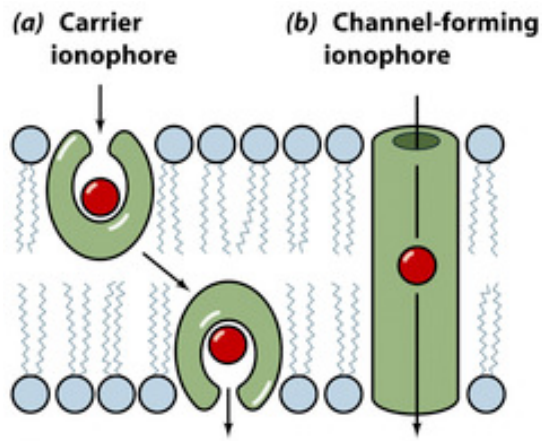
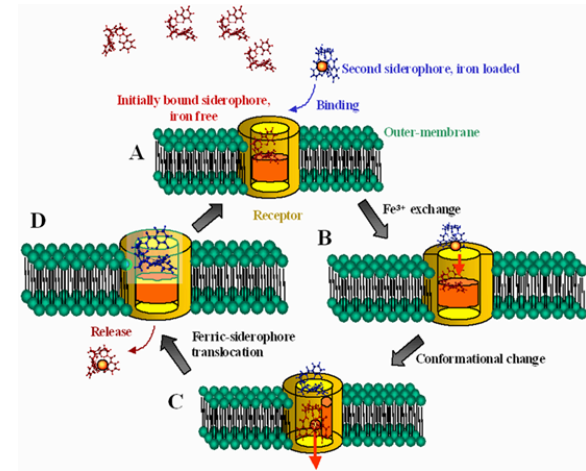
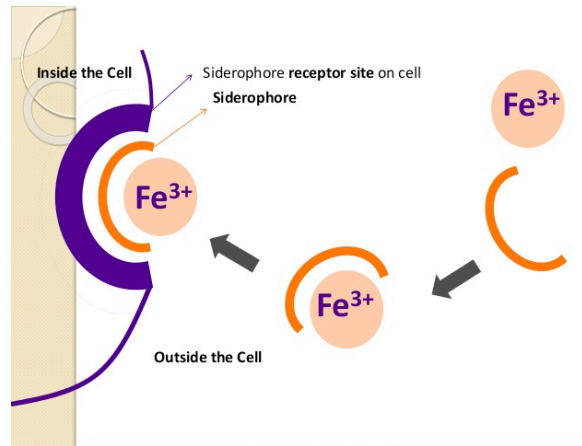


Fig. 5.12 Artificial ionophores and siderophores: (left to right) Na⁺-selective [15]crown-5, K⁺-selective [18]crown-6, Fe³⁺-selective cryptand and podand

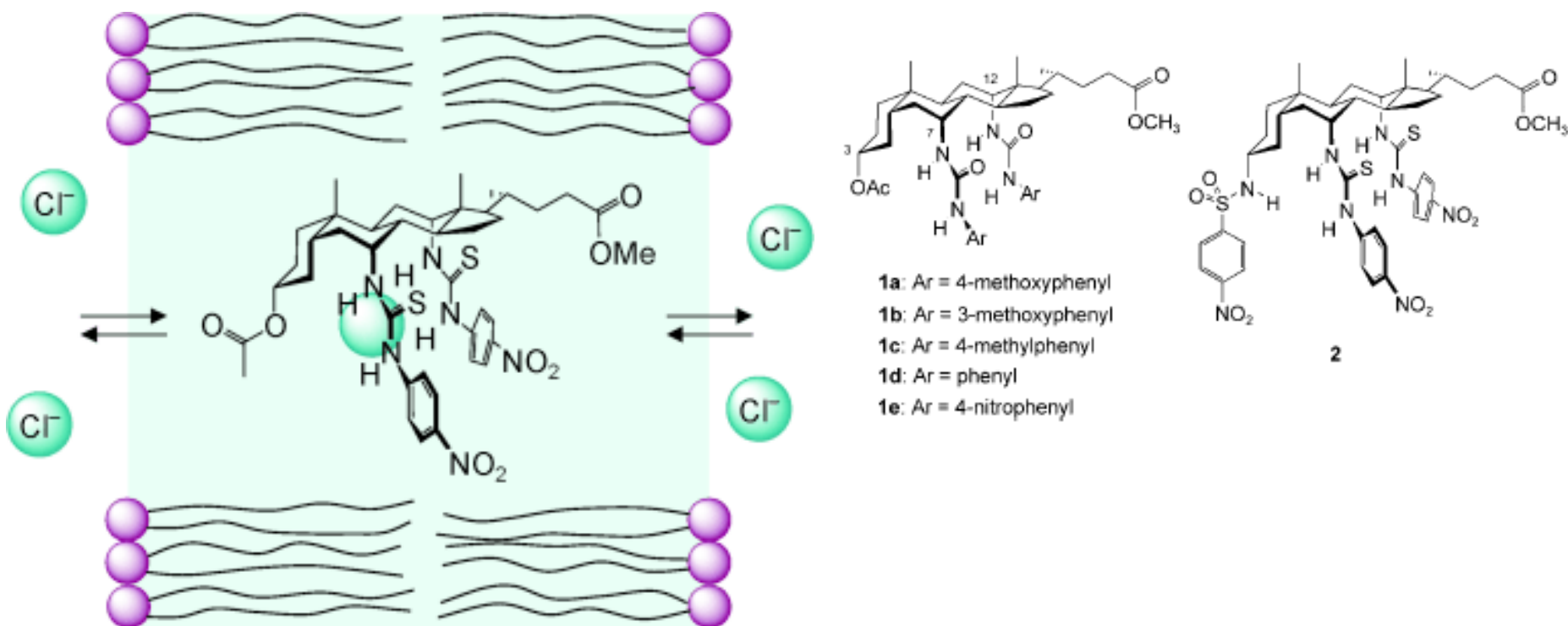


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Ken Raymond, Berkeley

Ion Transport: Anion Transport



McNally BA et al., *Chem Eur J*, **2008**, 14, 9599

Structure-activity relationships in cholapod anion carriers: enhanced transmembrane chloride transport through substituent tuning

Artificial Channel-Forming Peptides

Lear, Masserman, DeGrado, *Science*, 1988, 240, 1177

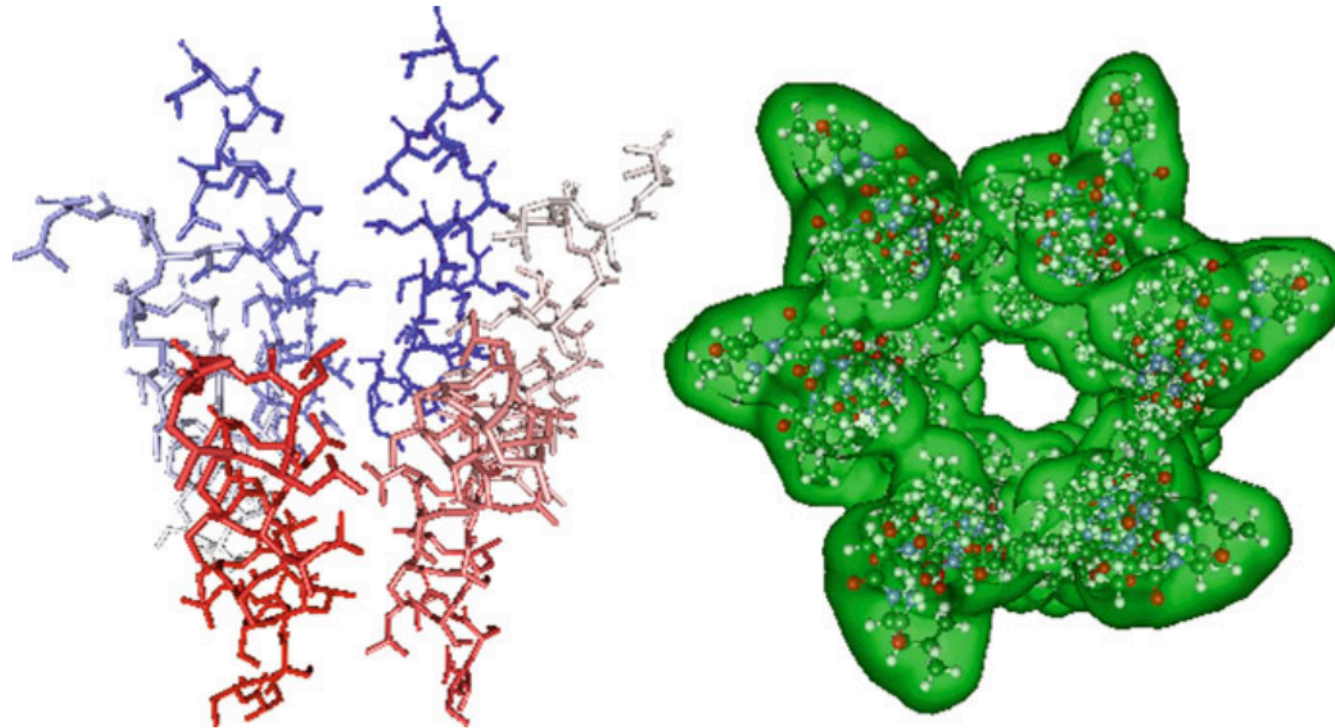


Fig. 5.13 Hexameric transmembrane channels formed by synthetic peptides [42]

A 21-residue peptide, $\text{H}_2\text{N}-(\text{Leu-Ser-Ser-Leu-Leu-Ser-Leu})^3-\text{CONH}_2$, which was designed to be a membrane-spanning amphiphilic α -helix, formed well-defined ion channels with ion permeability and lifetime characteristics resembling the acetylcholine receptor. In contrast, a 14-residue version of this peptide, which was too short to span the phospholipid bilayer as an α -helix, failed to form discrete, stable channels. A third peptide, $\text{H}_2\text{N}-(\text{ILu-Ser-Leu-Leu-Leu-Ser-Leu})_3-\text{CONH}_2$, in which one serine per heptad repeat was replaced by leucine, produced proton-selective channels. Computer graphics and energy minimization were used to create molecular models that were consistent with the observed properties of the channels.

Artificial Channel-Forming Peptides

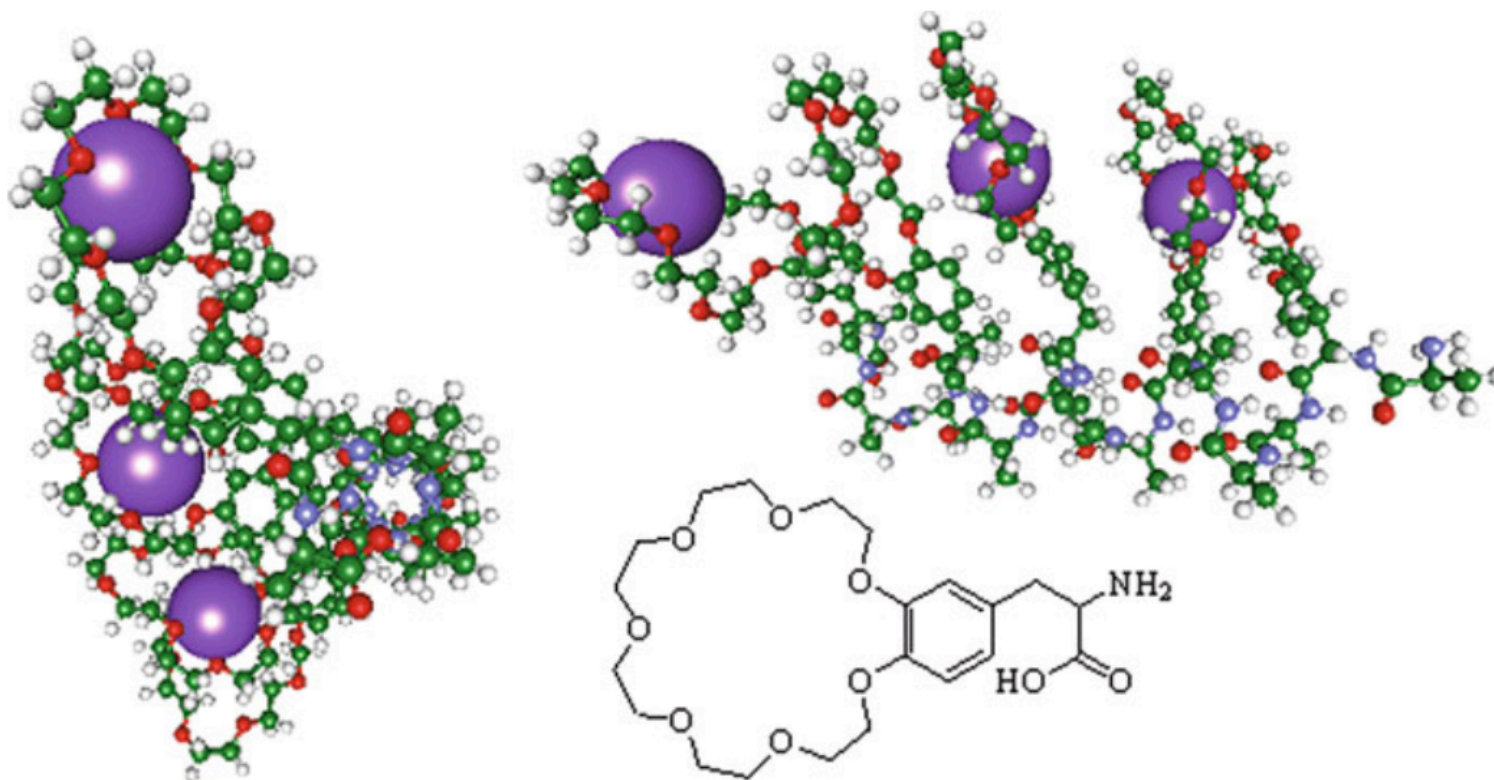


Fig. 5.14 Helical proteins incorporating a crown ether amino acid analogue: a view down the α -helix (*left*) and along the channel (*top*) [43]

Voyer N, Potvin L, Rousseau E, *J Chem Soc Perkin Trans*, **1997**, 2, 1469

Artificial Channel-Rigid Structures

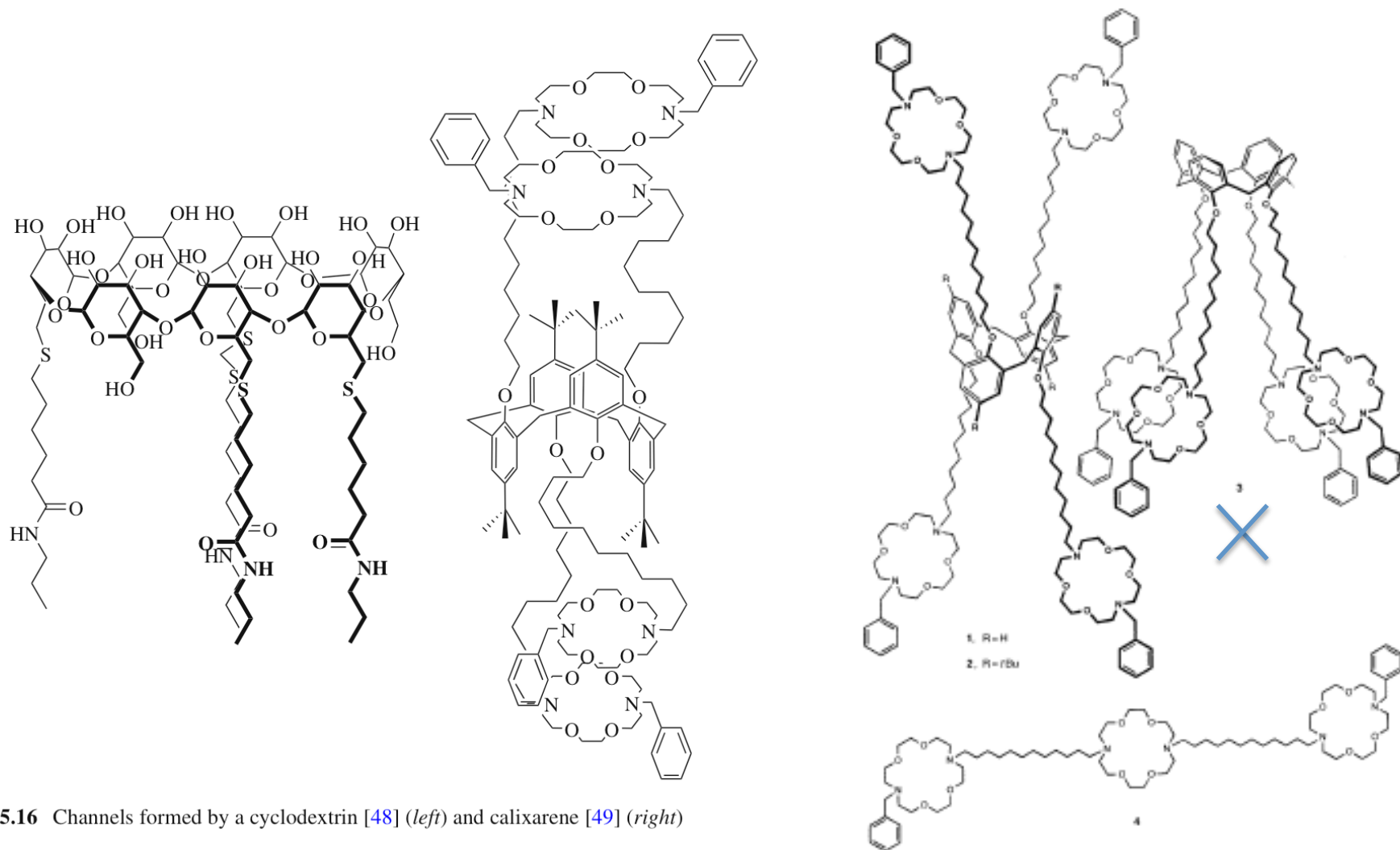
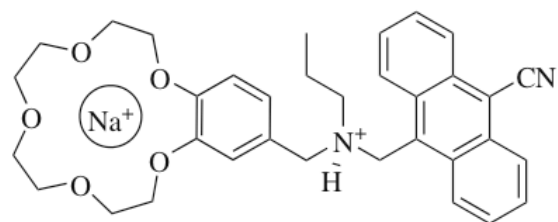


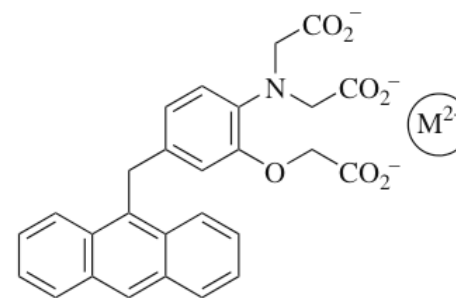
Fig. 5.16 Channels formed by a cyclodextrin [48] (left) and calixarene [49] (right)

Tabushi I, Kuroda Y, Yokata K. *Tetrahedron Let*, **1982** 23, 4601
de Mendoza J et al. *Angew Chem Int Ed*, **1998**, 37, 1534

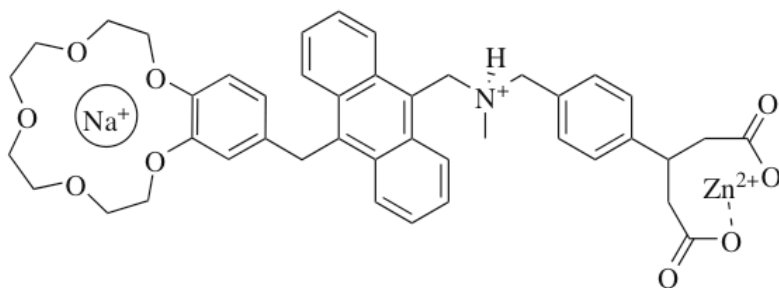
Diagnosis: Logic Gates



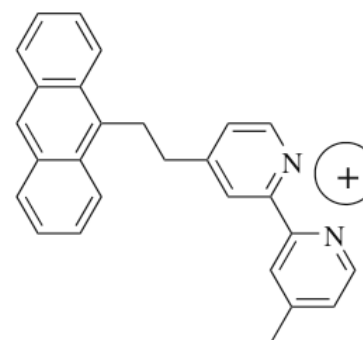
AND gate: $\text{Na}^+ + \text{H}^+ = \text{fluorescence}$



OR gate: any $\text{M}^{2+} = \text{fluorescence}$



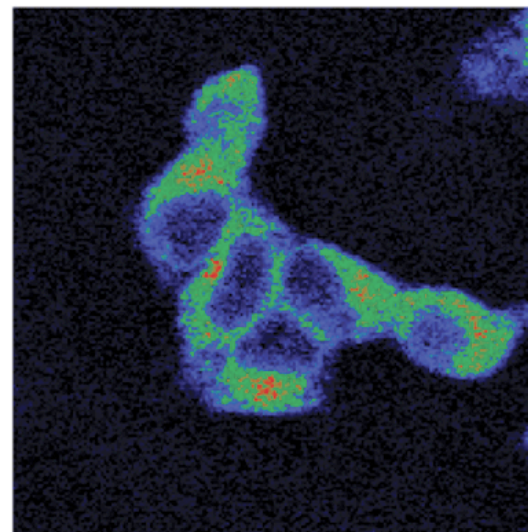
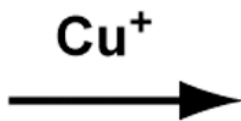
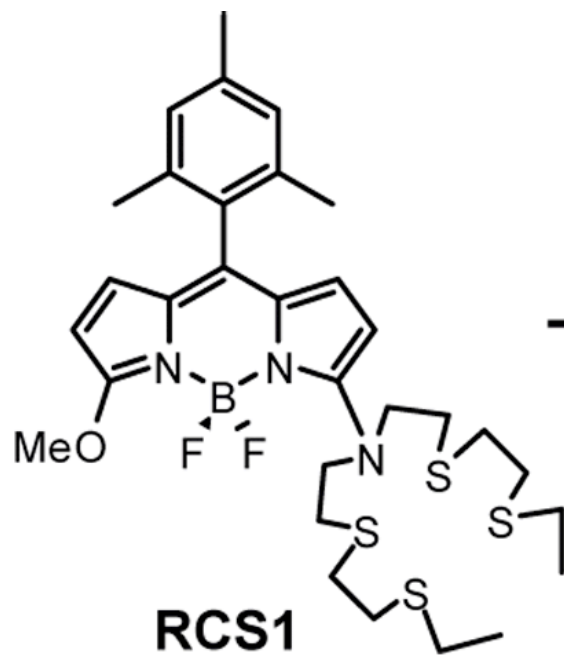
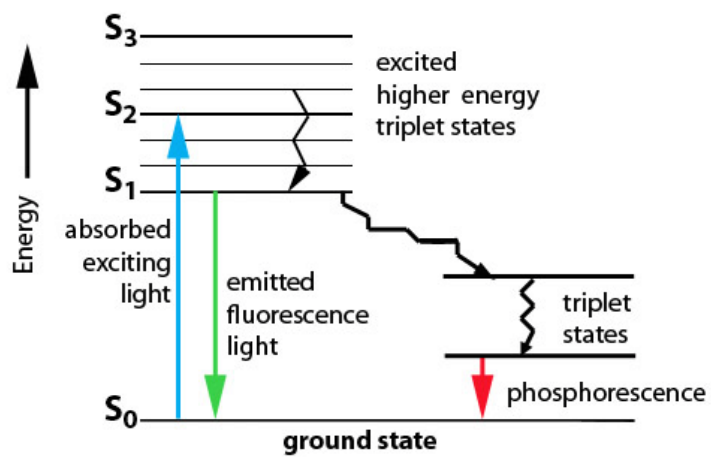
AND gate: $\text{Na}^+ + \text{H}^+ + \text{Zn}^{2+} = \text{fluorescence}$



NOR gate: fluorescence quenched by any guest

Fig. 6.5 Examples of logic gates

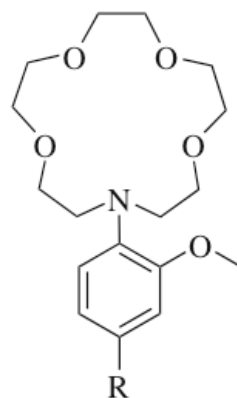
Diagnosis: Logic Gates



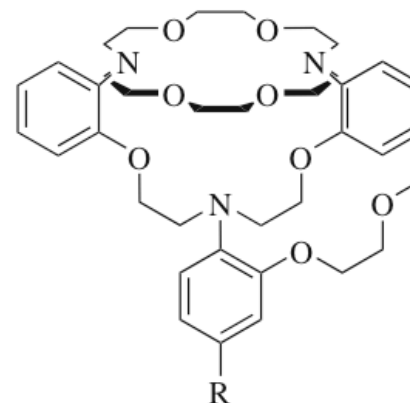
Diagnosis: Detection of critical analytes

The sensor gives an optical response and is therefore functions as an optode. In a similar approach, ionophore based sensors composed of cryptaspherands with chromogenic substituents have been developed to detect alkali metals in blood serum

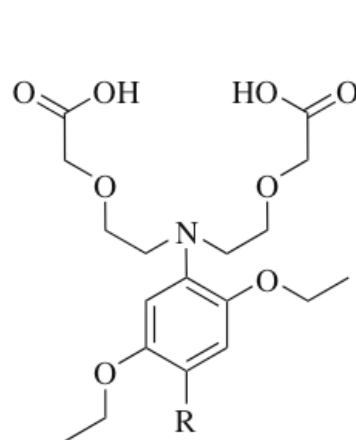
Kumar A et al (1988) Chromogenic ionophore-based methods for spectrophotometric assay of sodium and potassium in serum and plasma. Clin Chem 34:1709–1712



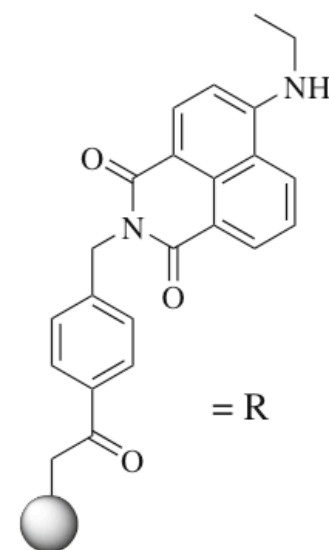
Na⁺ sensor



K⁺ sensor



Ca²⁺ sensor

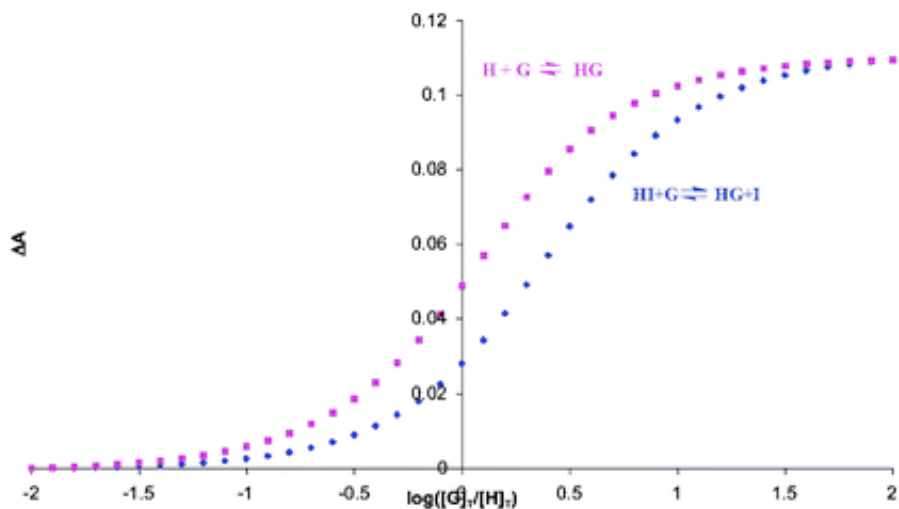
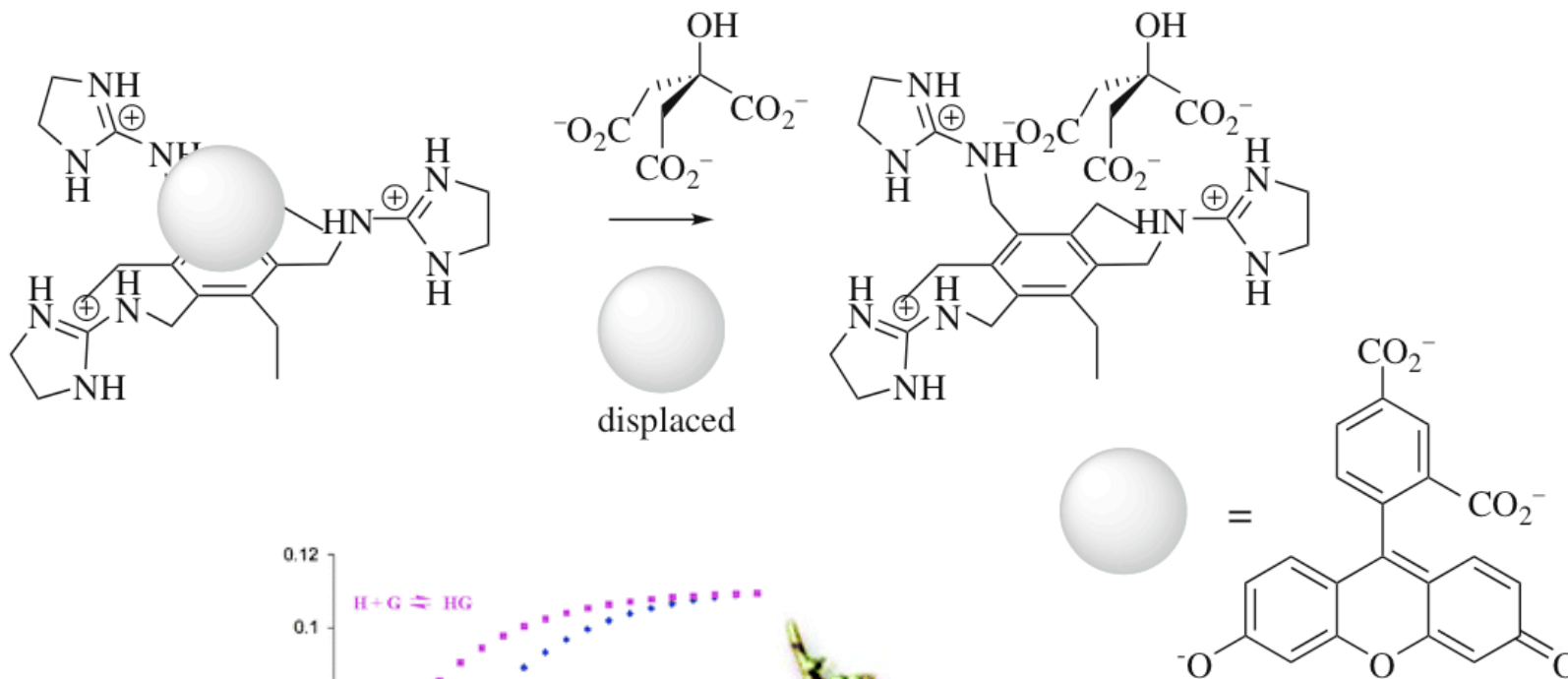


= R

Diagnosis: Displacement essay

Piatek AM et al, *J Am Chem Soc* 126, 6072

Threshold detection using indicator-displacement assays: An application in the analysis of malate in Pinot Noir grapes



Diagnosis: Squaraine Dyes

Gassensmith JJ et al, *J Am Chem Soc*, **2007**, *129*, 15054

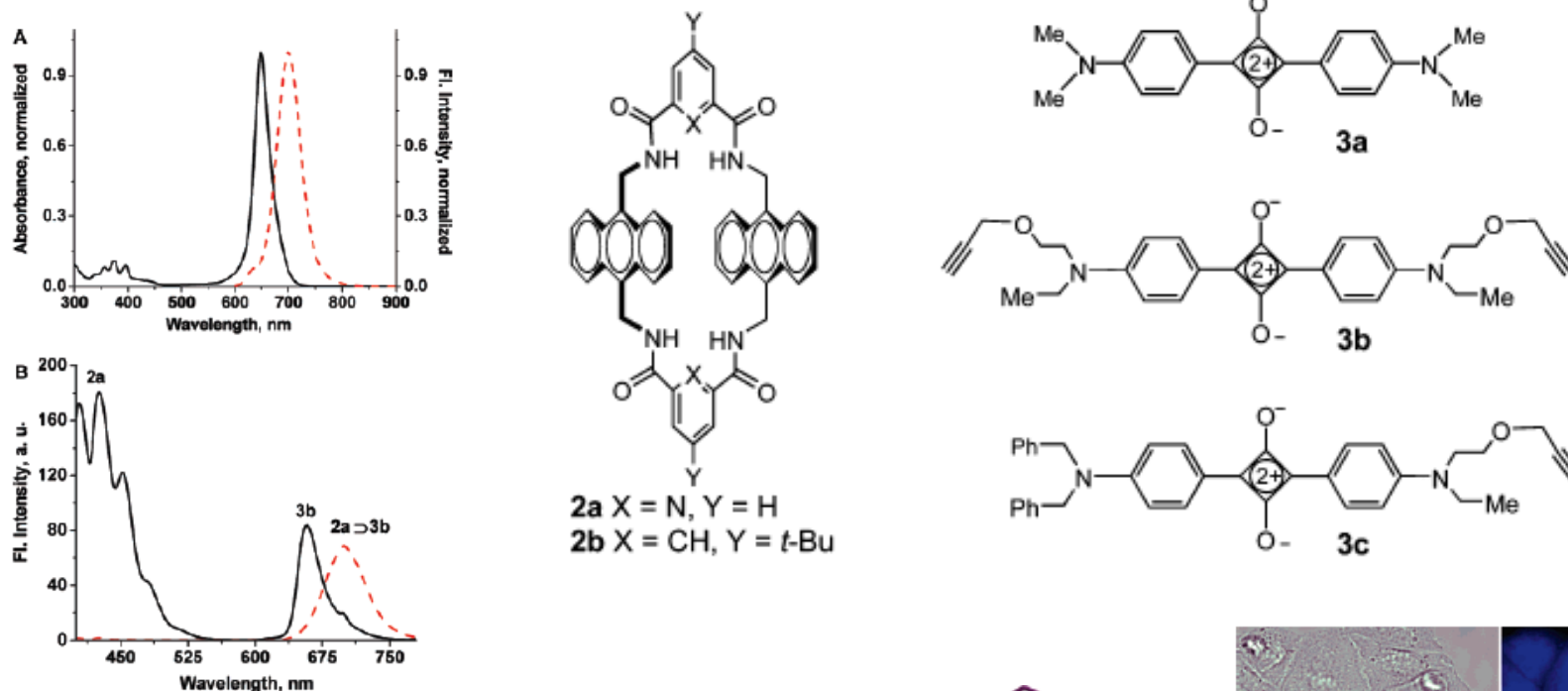


Figure 2. (A) Absorption (black full line) and fluorescence emission (red dashed line, ex: 580 nm) spectra for rotaxane 2a>3b in chloroform. (B) Fluorescence emission spectra (ex: 350 nm) for an equimolar mixture of 2a and 3b (both 5 μ M) (black full line) and a solution of 2a>3b (red dashed line) in chloroform.

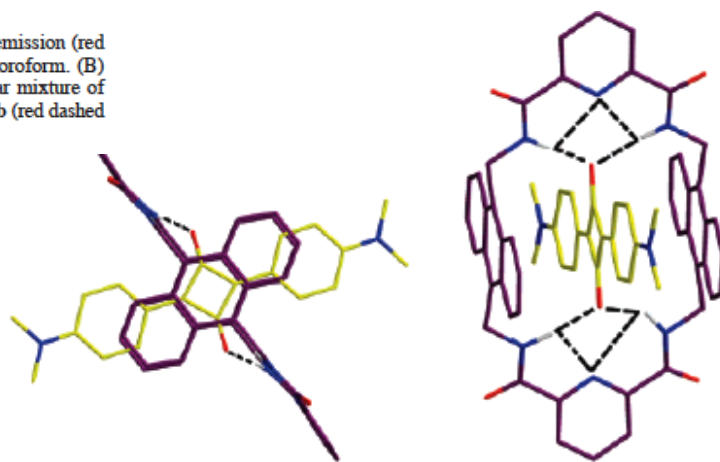


Figure 1. Two views of the X-ray crystal structure of 2a>3a.

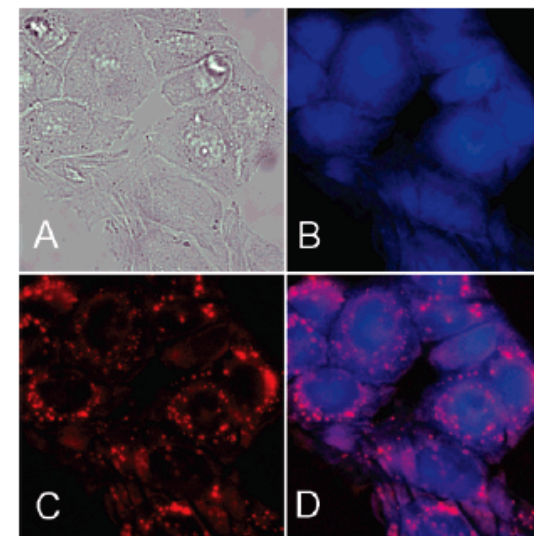
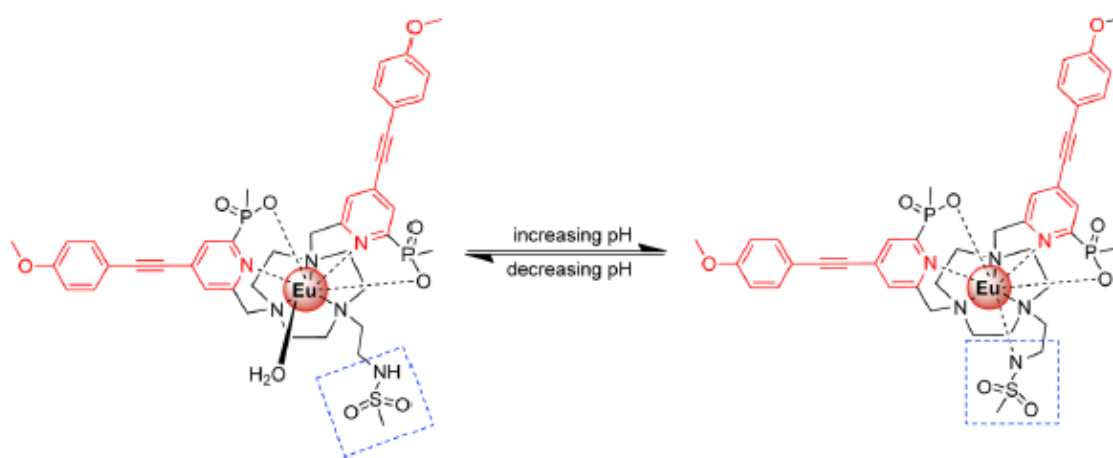
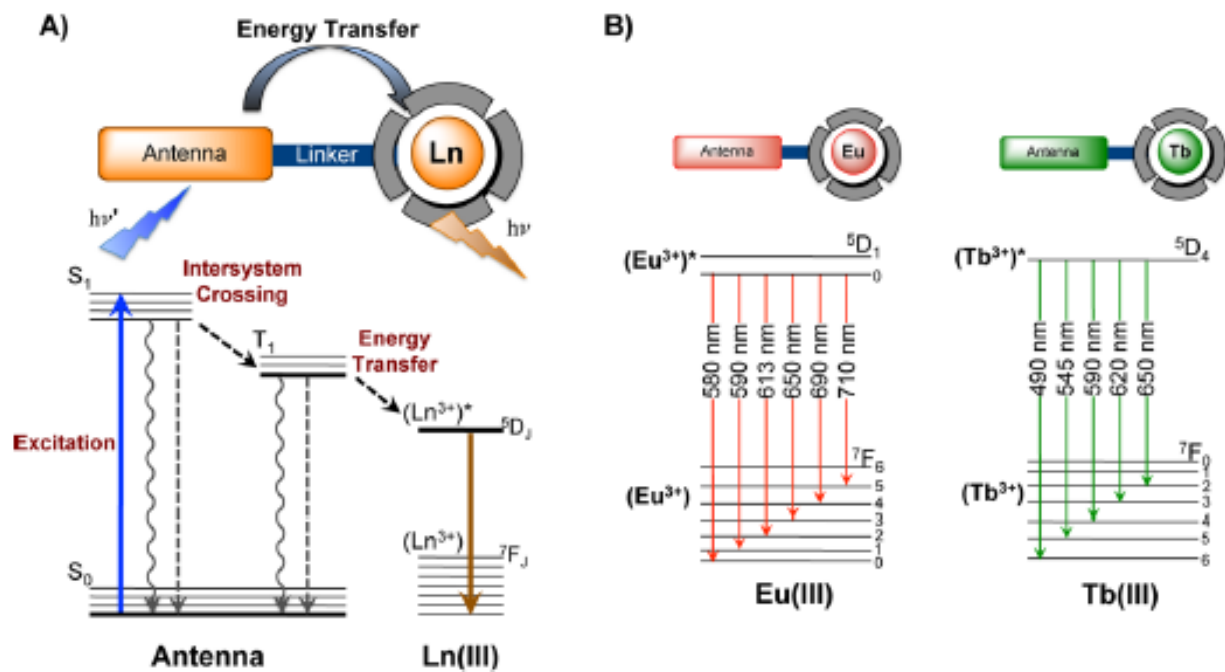


Figure 8. Fluorescence microscopy images of live CHO cells treated with separate aliquots of 3c (10 μ M) and 2b (10 μ M). Panel A: Phase contrast image. Panel B: Blue emission of uncomplexed 2b. Panel C: Far-red emission of 2b>3c. Panel D: Overlay of panels B and D.

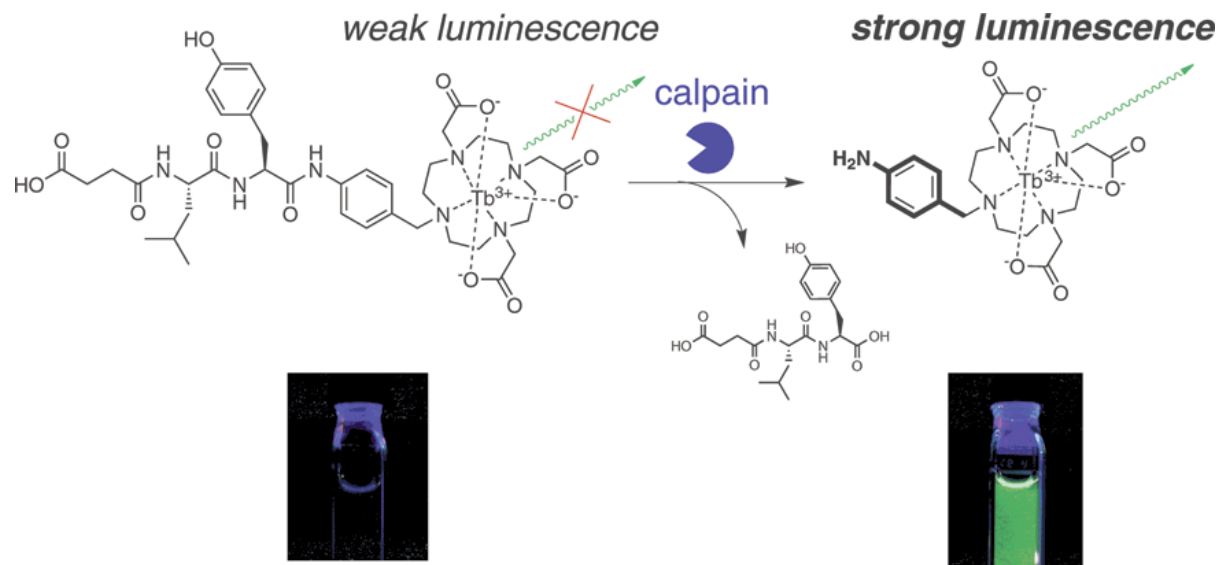
Diagnosis: In Vivo Imaging

Fluorescent agents: lanthanides

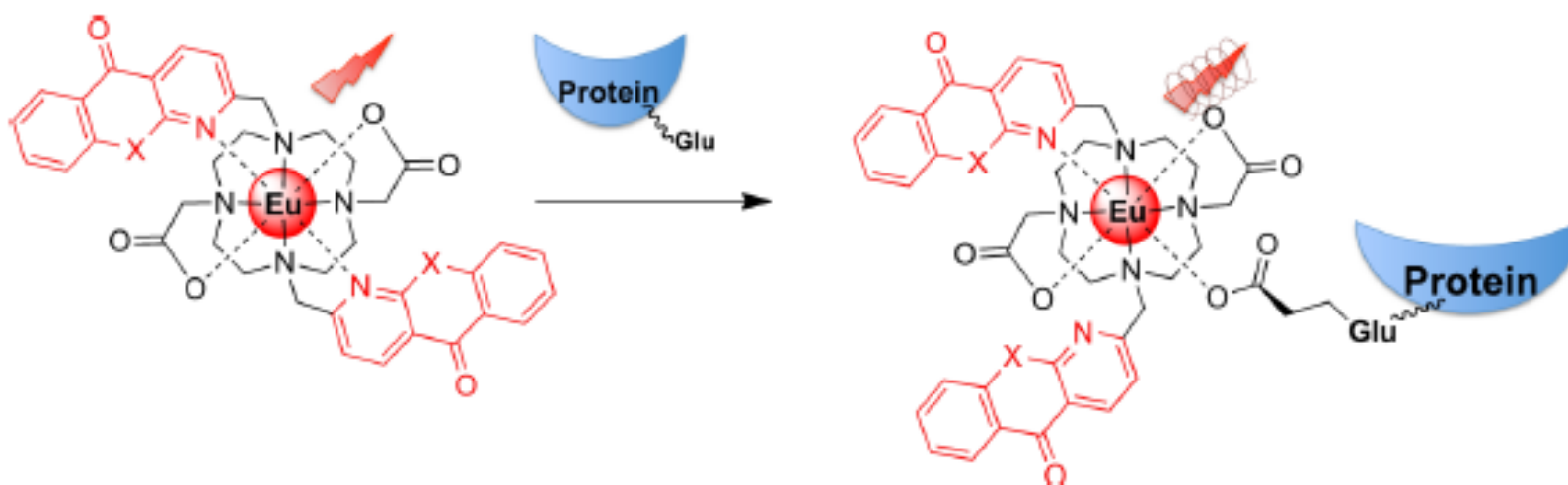


Diagnosis: In Vivo Imaging

Fluorescent agents



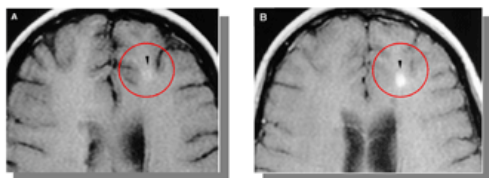
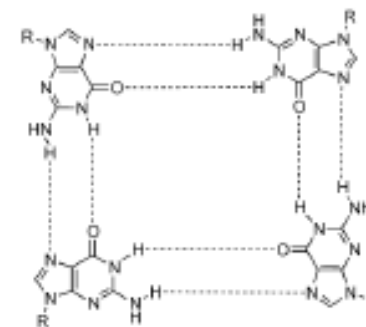
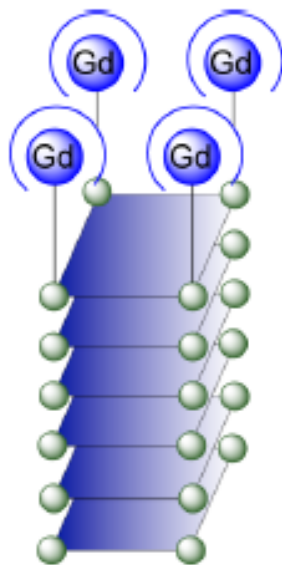
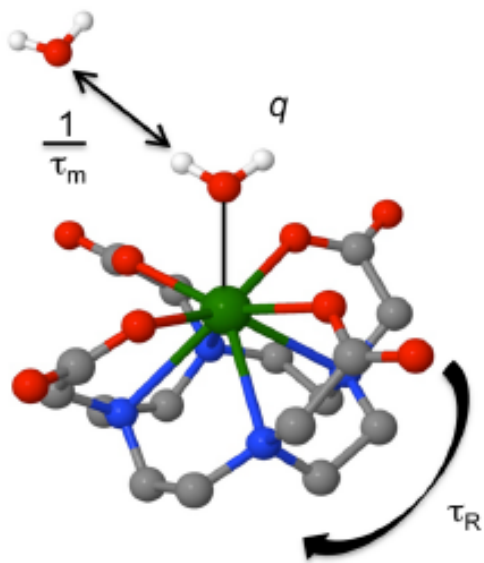
©2008 American Chemical Society *J. Am. Chem. Soc.* 130, 14376



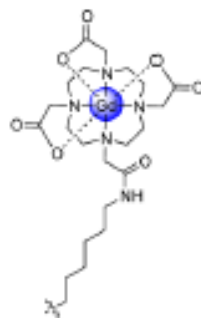
Peacock, *Dalton Trans.*, 2012, 41, 13154

Diagnosis: In Vivo Imaging

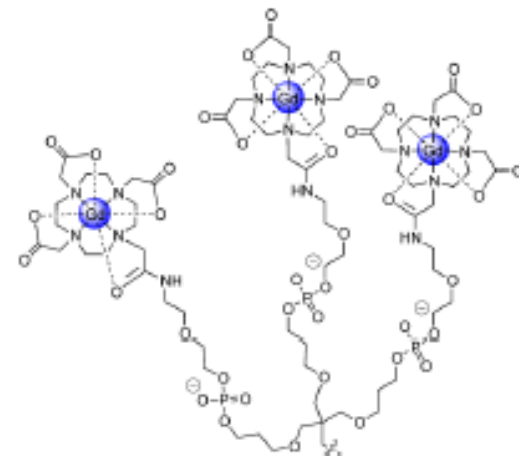
MRI Contrasts agents



Visualization of a tumor before (A) and after (B) injection of contrast agent.

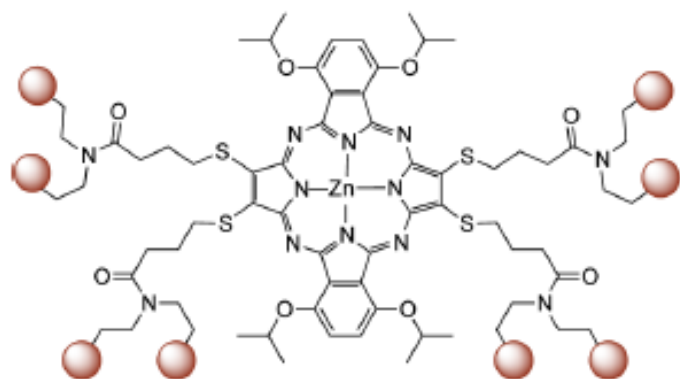


OR

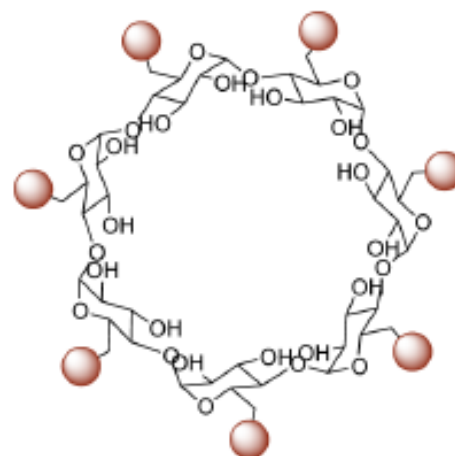


Diagnosis: In Vivo Imaging

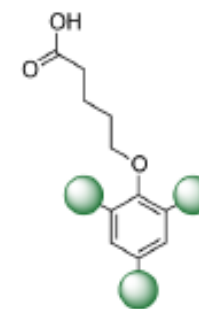
MRI Contrasts agents



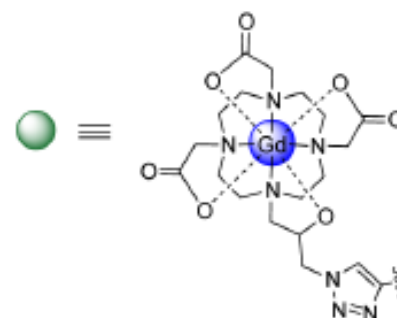
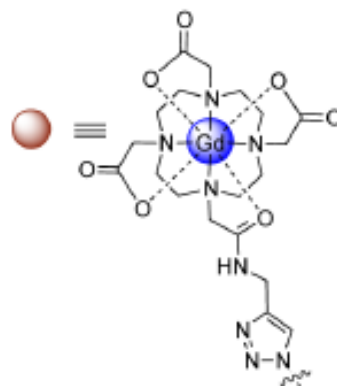
Zn-A₂B₂-8Gd(III)
 $r_1 = 12.8^A$



β -CD-Gd
 $r_1 = 12.2^B$

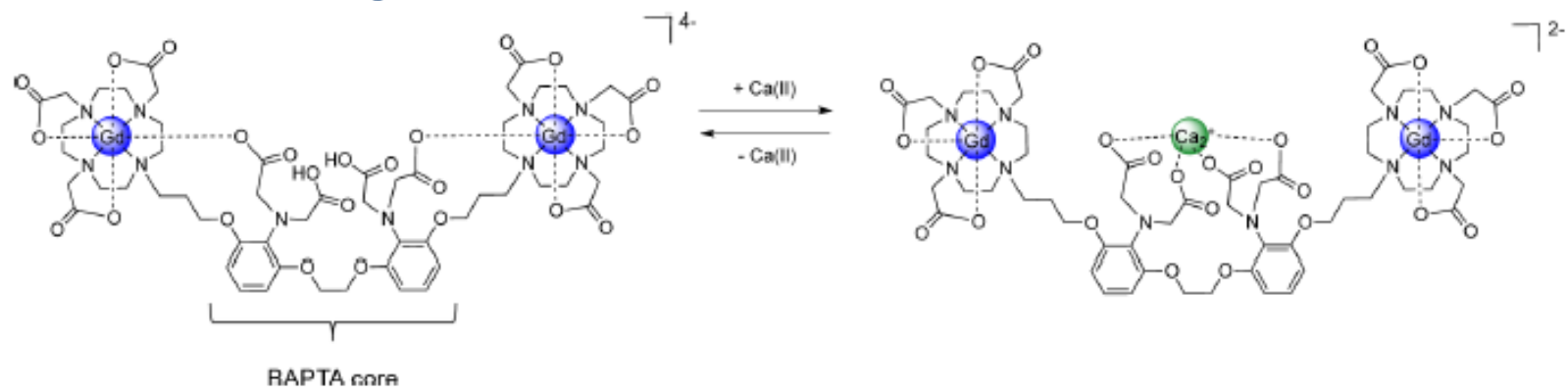


C3-Gd
 $r_1 = 15.4^C$

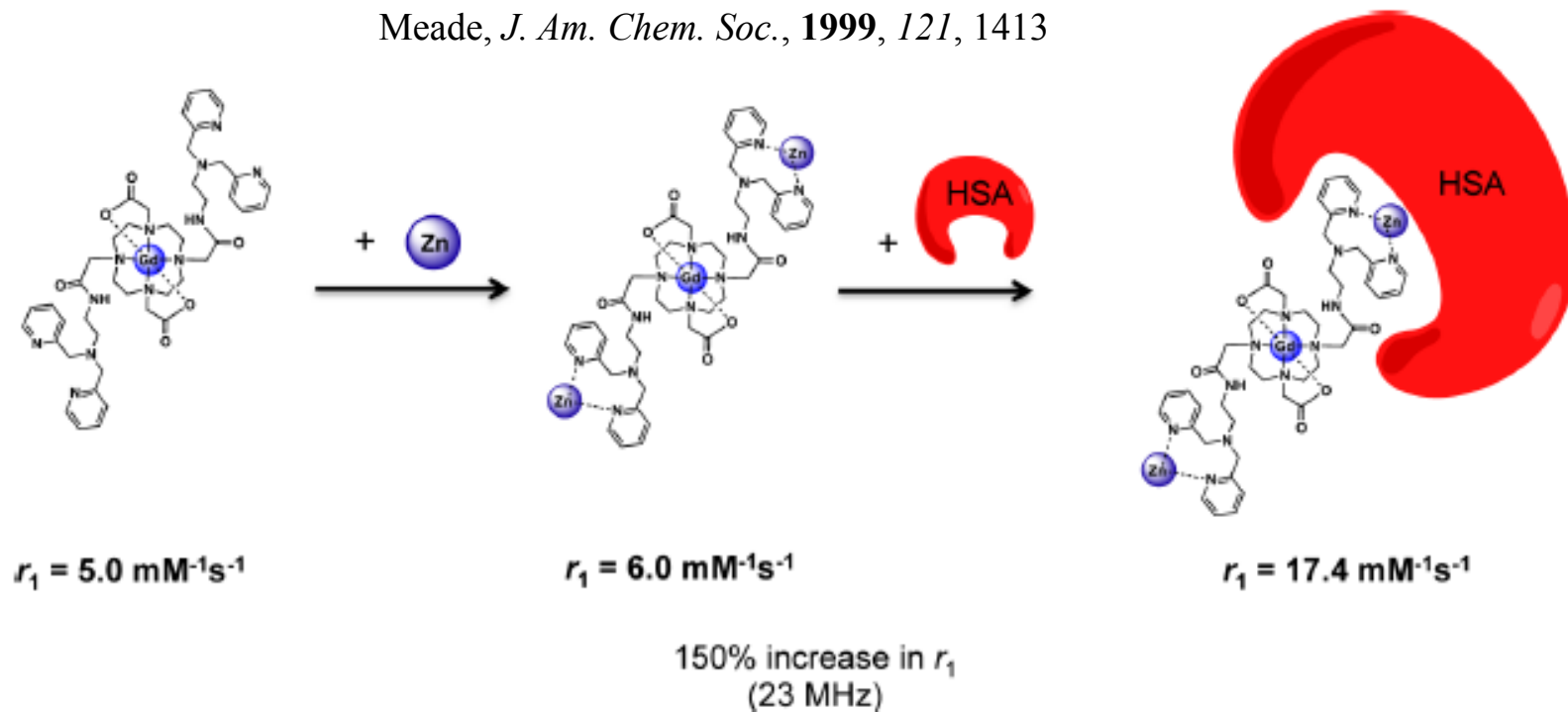


Diagnosis: In Vivo Imaging

MRI Contrasts agents



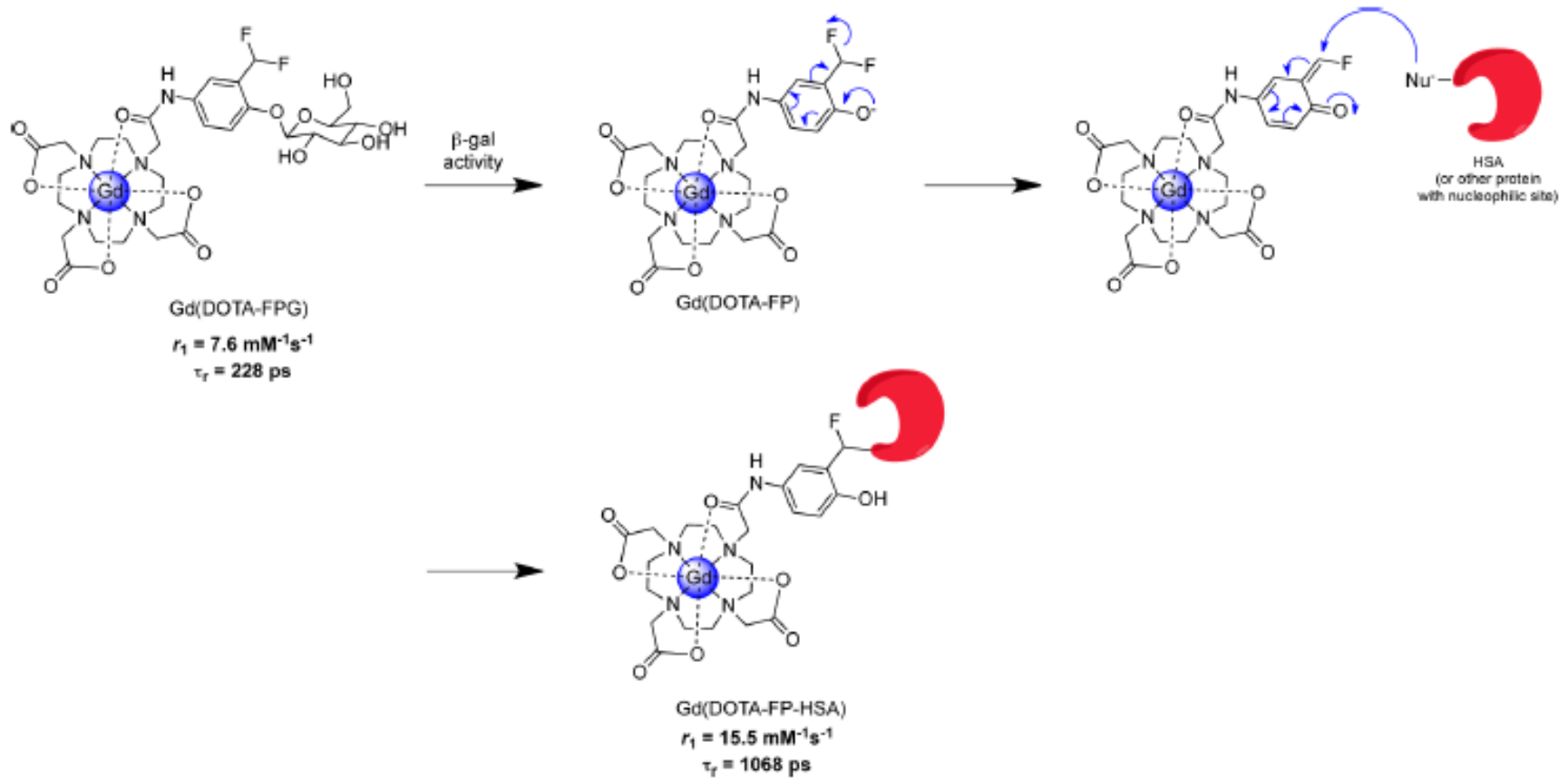
Meade, *J. Am. Chem. Soc.*, **1999**, *121*, 1413



Sherry, *J. Am. Chem. Soc.*, **2009**, *131*, 11387

Diagnosis: In Vivo Imaging

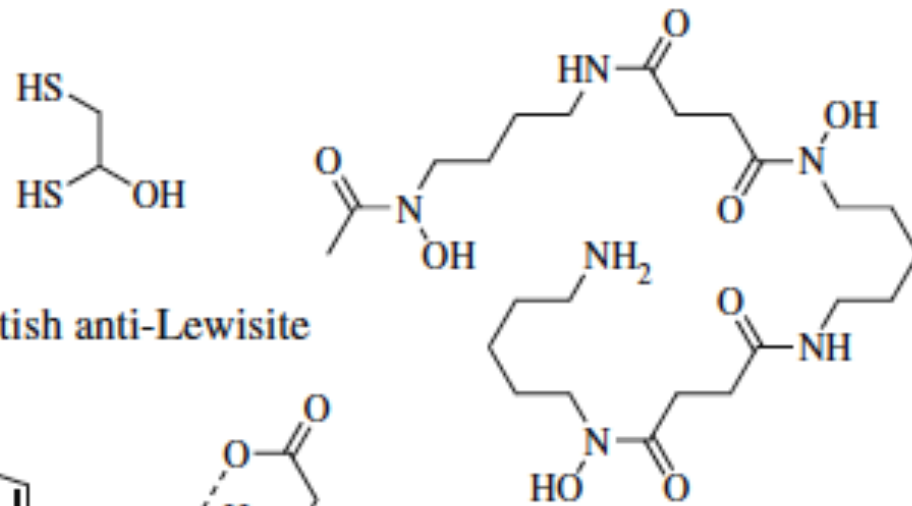
MRI Contrasts agents



Chang, *Bioconjugate Chem.*, **2007**, *18*, 1716

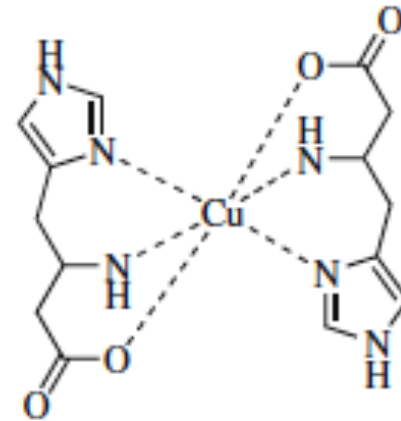
Supramolecular Therapeutics

Chelation therapy

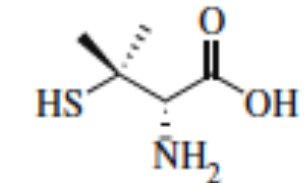


British anti-Lewisite

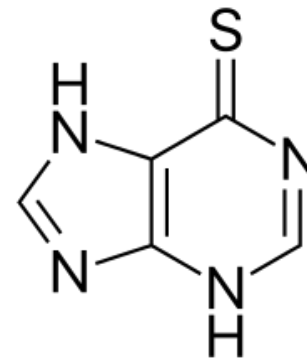
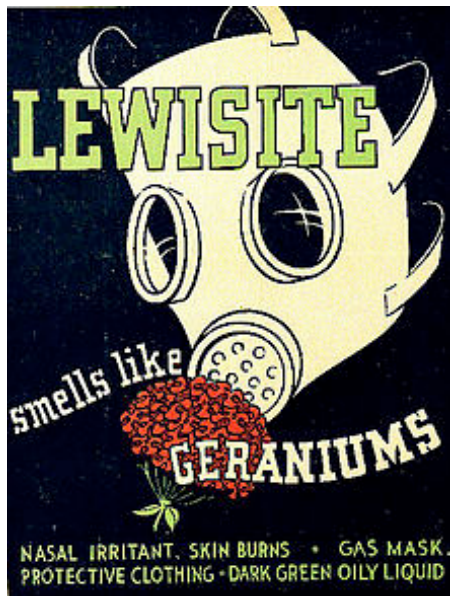
desferrioxamine



copper histidine complex



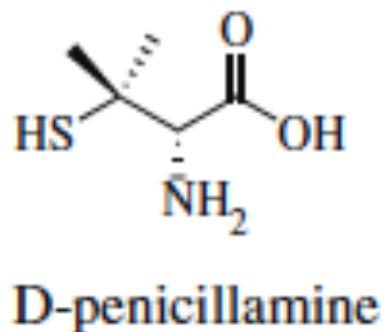
D-penicillamine



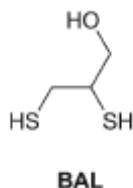
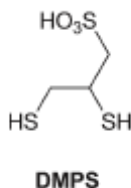
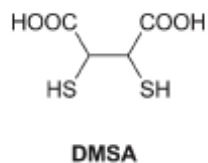
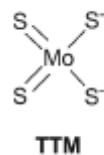
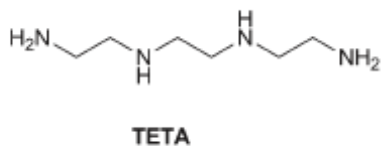
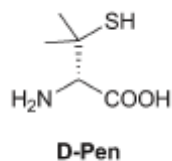
Mercaptopurine

Copper Imbalance: Wilson diseases

Chelation therapy

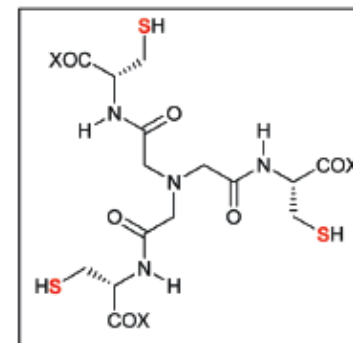


Wilson's disease or hepatolenticular degeneration is an autosomal recessive genetic disorder in which copper accumulates in tissues; this manifests as neurological or psychiatric symptoms and liver disease. It is treated with medication that reduces copper absorption or removes the excess copper from the body, but occasionally a liver transplant is required.



Tripodal pseudopeptides

- H₃L¹: X = OEt
- H₃L²: X = NH₂
- H₆L³: X = OH

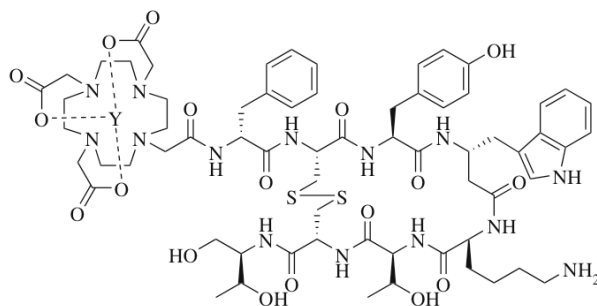


Delangle, Dalton Trans., **2012**, *41*, 6359

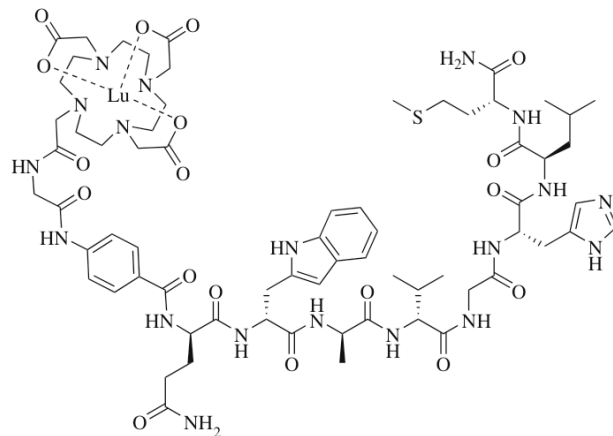
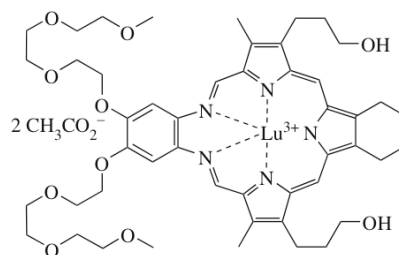
Supramolecular Therapeutics

Macrocyclic Complexes for Radiotherapy

^{90}Y , half time of 64h



Photodynamic Therapy



porphyria

Fig. 7.3 Radiotherapeutics: (from *top*) DOTA—tyr3-octreotide, LUTRIN[®] and Lu-177-AMDA

Supramolecular Therapeutics

Photodynamic Therapy

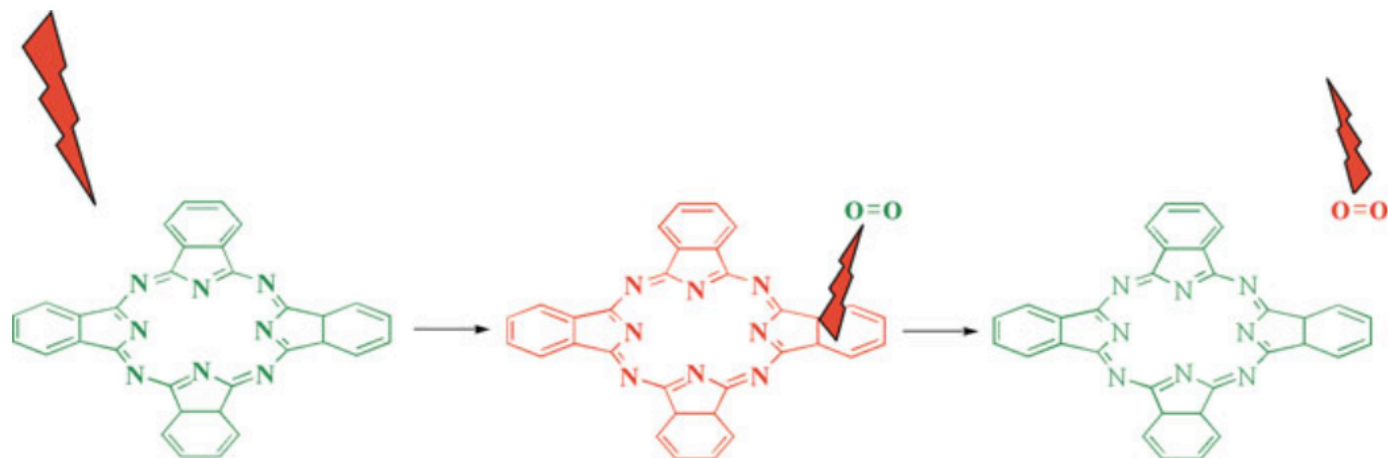


Fig. 7.4 Generation of therapeutic $^1\text{O}_2$ for photodynamic therapy

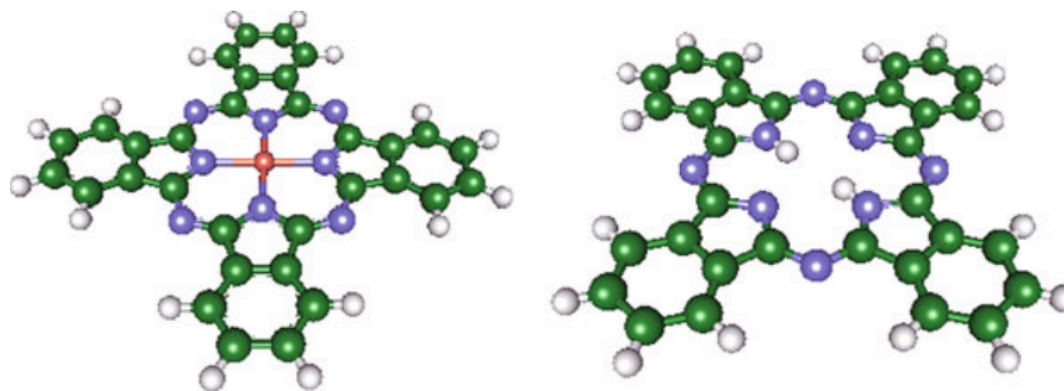
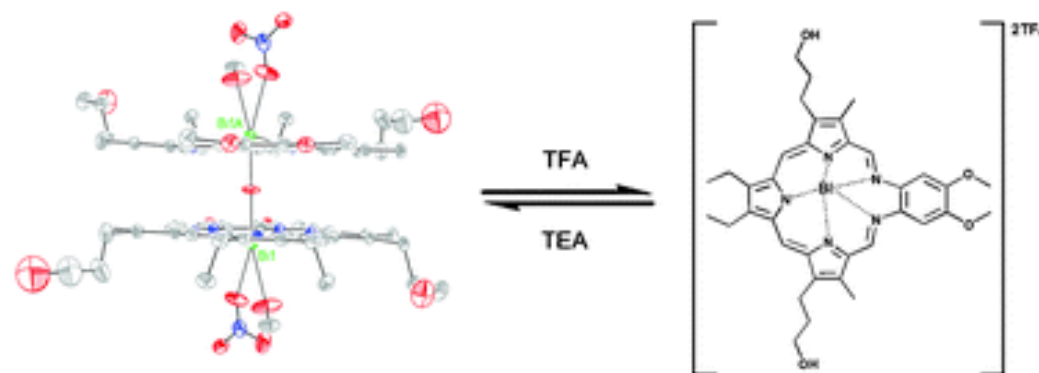
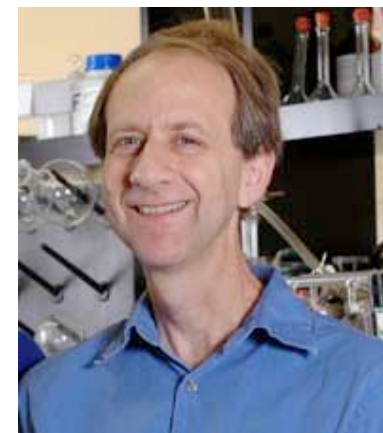
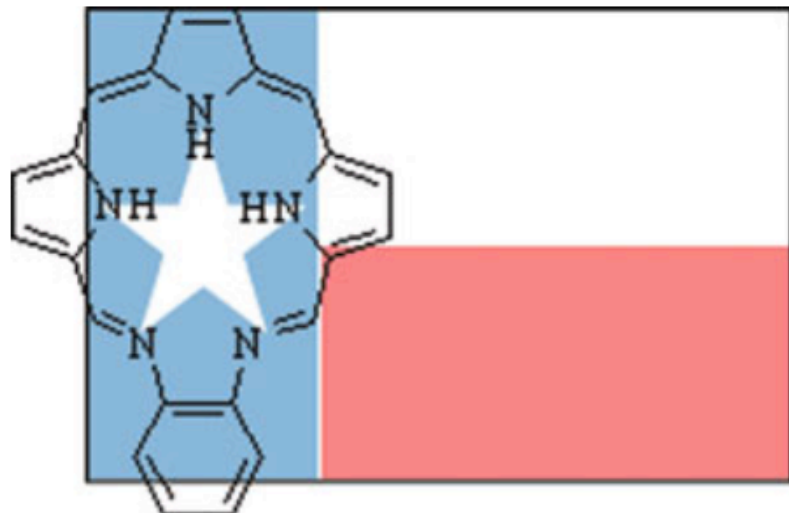


Fig. 7.5 Phthalocyanine crystal structures: a copper complex [12] (*left*) and the metal free macrocycle [13] (*right*)

Supramolecular Therapeutics

Texaphyrin



Sessler, *Chem. Commun.*, **2010**, 46, 7900

Sessler JL et al., Synthesis and structural characterization of lanthanide(III) texaphyrins., 1993, *Inorg Chem* 32, 3175

Sessler JL, Miller RA, Texaphyrins – New drugs with diverse clinical applications in radiation and photodynamic therapy. *Biochem Pharmacol* , 2002, 59:733

Drug Delivery and Controlled Release

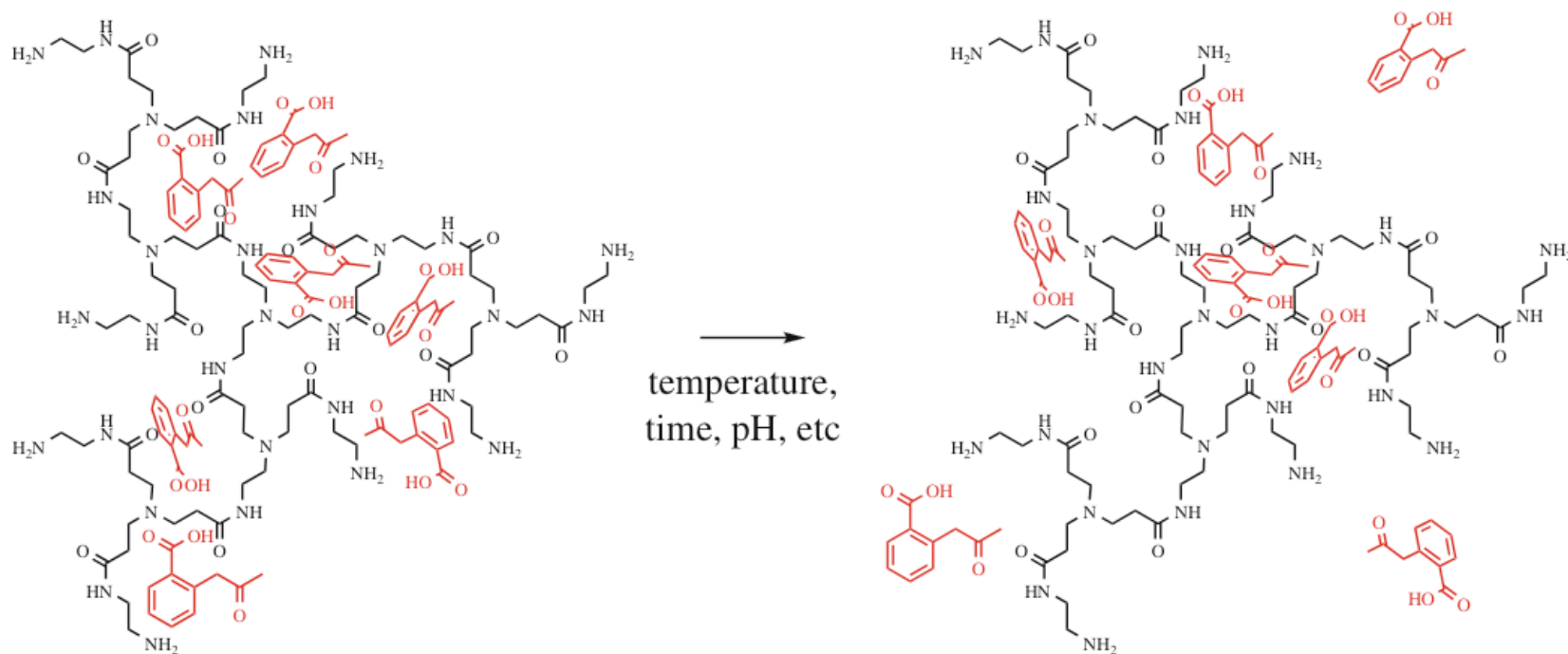


Fig. 7.8 Controlled release of therapeutic drugs by a dendrimer

Malik N, Evagorou EG, Duncan R (1999) Dendrimer-platinate: a novel approach to cancer chemotherapy. *Anti-Cancer Drugs* 10: 767–776

Padilla De Jesus OL et al (2002) Polyester dendritic systems for drug delivery applications: *in vitro* and *in vivo* evaluation. *Bioconjug Chem* 13:453–461

Cyclams as Anti-HIV Agents

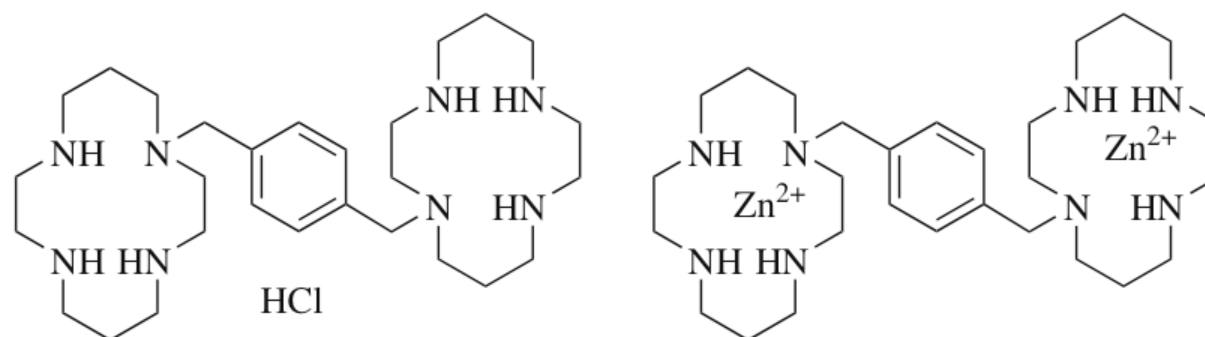
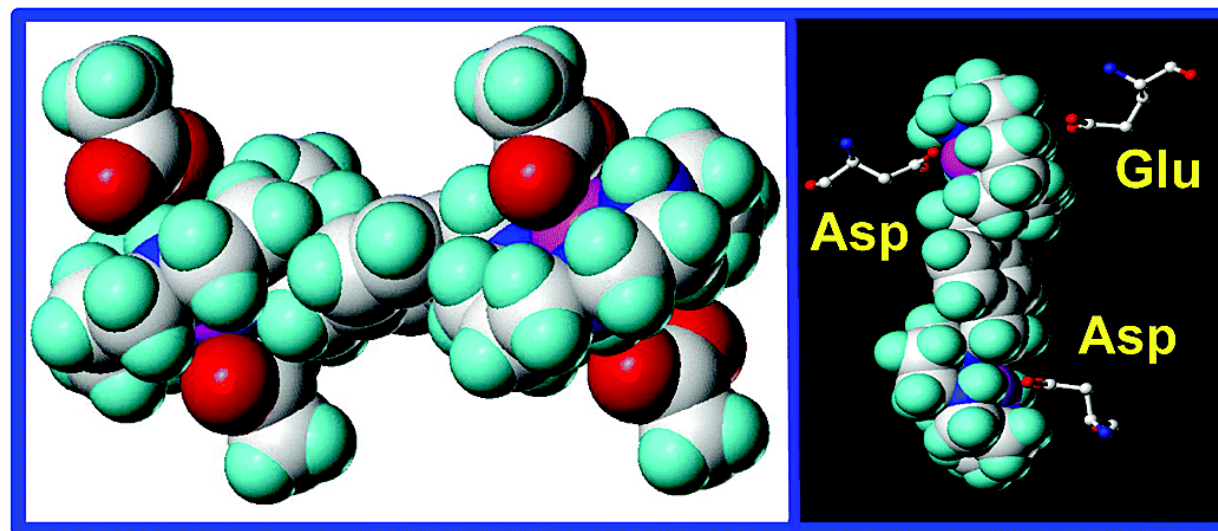


Fig. 7.9 Bis(cyclam)s with anti-HIV activity

Structure and dynamics of metallomacrocycles, Recognition of zinc xylyl-bicyclam by an HIV coreceptor.



A Supramolecular Solution to Alzheimer's Disease?

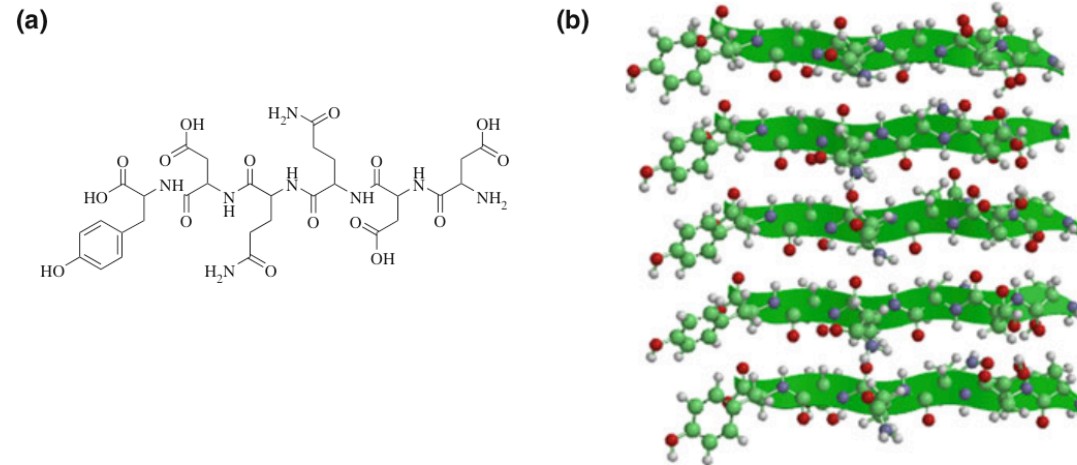
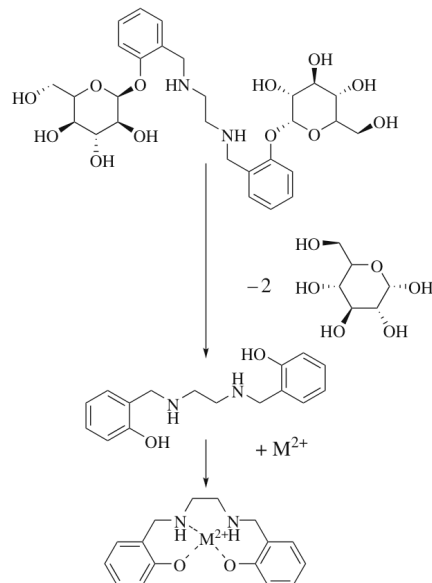


Fig. 7.10 The β -sheet structure formed by the NNQQNY sequence (a) in amyloid- β proteins that leads to plaque formation [30]

Fig. 7.11 A tetrahydrosalen-based therapy to target Alzheimer's disease [31]



Goldschmidt L et al (2010) Identifying the amyloids, proteins capable of forming amyloid-like fibrils. *Proc Natl Acad Sci USA* 107:3487–3492

Stor T et al (2009) Glycosylated tetrahydrosalens as multifunctional molecules for Alzheimer's therapy. *Dalton Trans* 3034–3043

Calixarenes as Therapeutic Agents

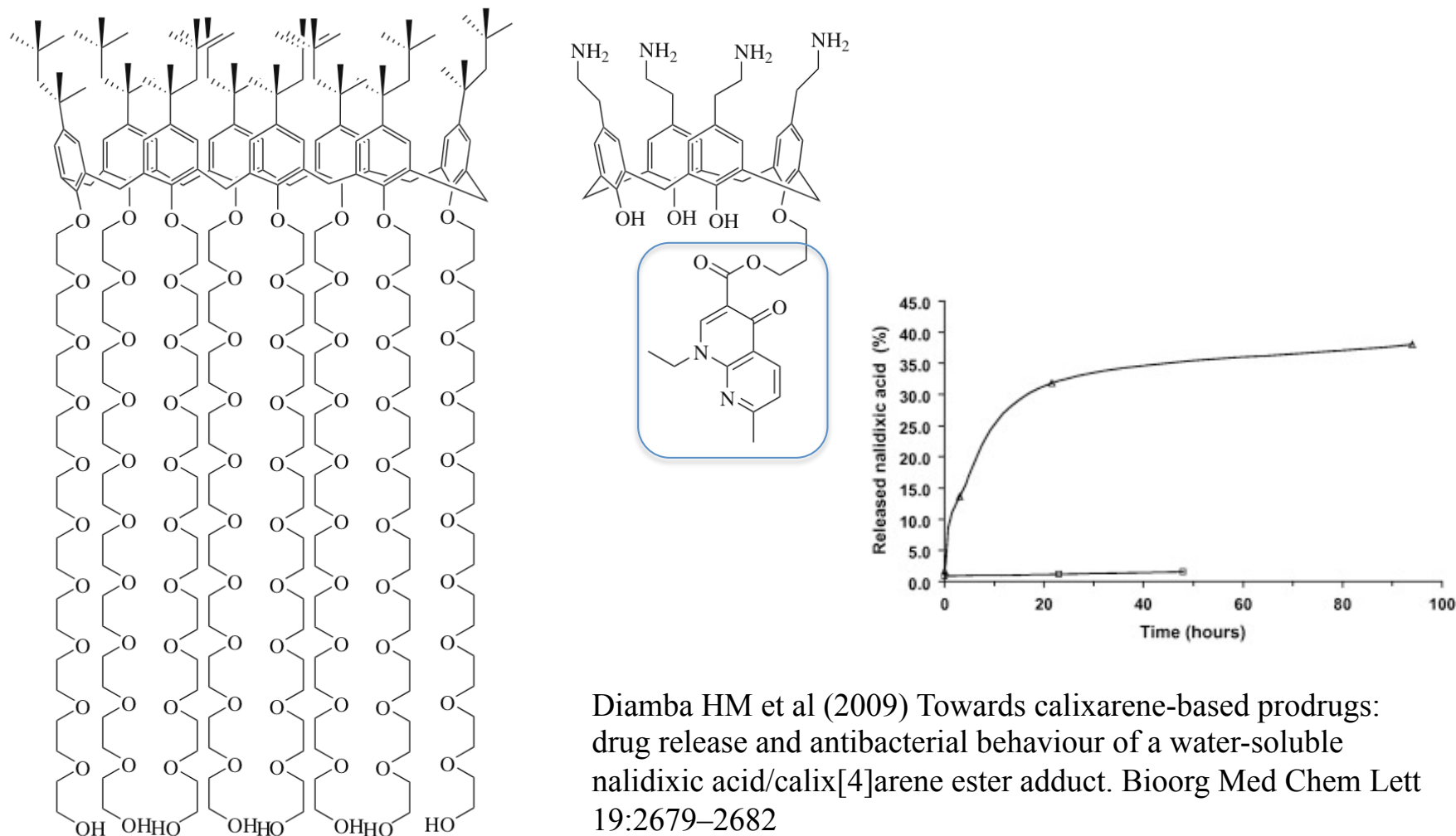


Fig. 7.12 Calixarene-based therapeutics: anti-tubercular *Macrocyclon* [32] (left) and a naldixic acid delivering prodrug [34] (right)

Antitubercular agent, 1955

Supramolecular Antibiotics

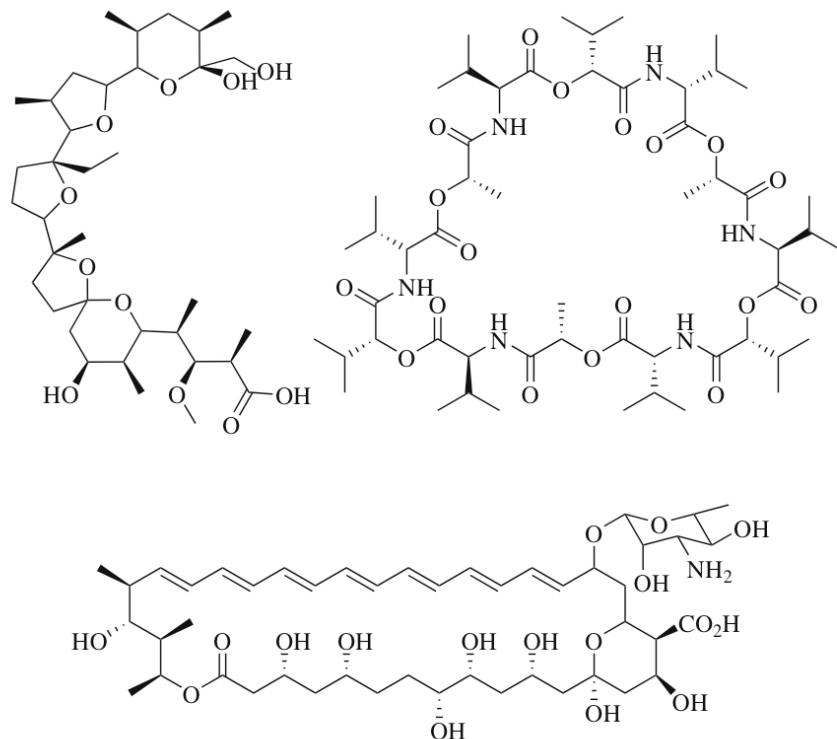


Fig. 7.13 Some naturally occurring antibiotics and related compounds: monensin (*top left*), valinomycin (*top right*) and amphotericin (*bottom*)

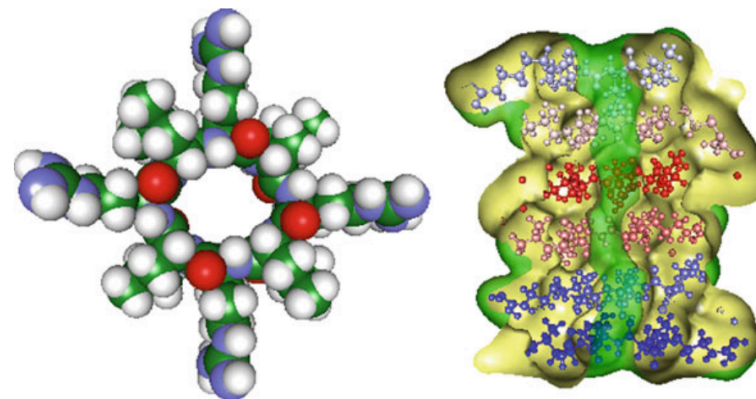


Fig. 7.14 Self-stacking cyclopeptides [36]

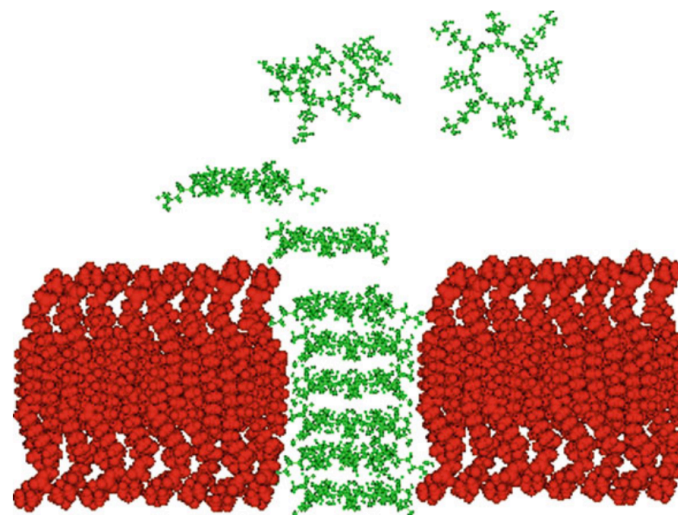


Fig. 7.15 Proposed mode of antibiotic action by cyclopeptides: tubes (*green*) form and aggregate to disrupt bacterial membranes (*red*)

Fernandez-Lopez S et al (2001) Antibacterial agents based on the cyclic D,L- α -peptide architecture. *Nature* 412:452–455

Ruzin A et al (2004) Mechanism of action of the mannopeptimycins, a novel class of glycopeptide antibiotics active against vancomycin-resistant gram-positive bacteria. *Antimicrob Agents Ch* 48:728–738

Motiei L et al (2009) Antibacterial cyclic D,L- α -glycopeptides. *Chem Commun* 3693–3695